BIOGENIC CARBON FRACTION OF BIOGAS AND NATURAL GAS FUEL MIXTURES DETERMINED WITH ¹⁴C

DOI: 10.2458/56.16514

Sanne W L Palstra^{1,2} • Harro A J Meijer¹

ABSTRACT. This study investigates the accuracy of the radiocarbon-based calculation of the biogenic carbon fraction for different biogas and biofossil gas mixtures. The focus is on the uncertainty in the 14 C reference values for 100% biogenic carbon and on the 13 C-based isotope fractionation correction of the measured 14 C values. The separately (AMS) measured CO 2 and CO 4 fractions of 8 different biogas samples showed 14 C values between 102% and 116% (pMC). The CO 5 values of these samples varied between $^{-6}$ 6% and $^{+3}$ 1% for the CO 2 fraction and between $^{-28}$ 8% and $^{-62}$ 9% for the CO 4 fraction. The uncertainty in calculated biogenic carbon fractions due to uncertainty in the 14 C reference values depends on the available information about the origin of the used biogenic materials. It varies between $^{\pm0}$ 5% and $^{\pm3}$ 5% (absolute) depending on the type of biogas. A method is proposed to minimize this uncertainty for different groups of biogases. The calculated biogenic carbon fraction deviates up to $^{\pm2}$ 5% for biofossil gas mixtures, if the applied isotope fractionation correction is based on the 51 6 Value of the mixed biofossil sample instead of the biogenic 51 6 Value. Combination of both error sources shows that the uncertainty in the calculated biogenic carbon fraction varies between $^{\pm0}$ 7% and $^{\pm4}$ 5%, depending on the type of biogas in the sample.

INTRODUCTION

Over the last 10–15 yr, several international policy measures have been introduced aiming at reducing the dependence on fossil fuels and decreasing the amount of fossil fuel CO_2 emissions (European Commission 2003). The use of biomass in different fuel products has been encouraged by many financial incentives. Due to the large financial interests in the production, trading, and use of biomass-based fuels and/or their CO_2 emissions, reliable verification methods are needed to distinguish biogenic from fossil carbon fractions in fuels and in CO_2 emissions (flue gas).

Arguably the most reliable verification method so far for fuels and flue gas CO₂ is the ¹⁴C method. In this method, the measurement of the radioactive carbon isotope ¹⁴C in a fuel or flue gas CO₂ sample is used to determine the biogenic and fossil carbon fractions of the sample (in short: bioC fraction and fossilC fraction, respectively). The method is based on the linear relation between the measured ¹⁴C abundance in a sample and the dilution rate of biogenic ¹⁴C in the sample due to the fossil ¹⁴C-free carbon fraction in the sample. The accuracy of the ¹⁴C method to determine the bioC fraction of a specific sample (batch) depends on (1) representative carbon sampling and sample pretreatment, (2) accurate ¹⁴C measurement, and (3) correct calculation of the bioC fraction.

So far, the ¹⁴C method has been investigated and tested for different liquid and solid fuels and for flue gas CO₂ from the combustion of different fuel materials at different industrial/energy plants (Dijs et al. 2006; Mohn et al. 2008; Staber et al. 2008; Palstra and Meijer 2010). The ¹⁴C method has not been demonstrated yet for fuel mixtures containing biogas and natural gas.

The use of biogas (specifically, its biomethane fraction) as an alternative for natural gas increases due to more demand and an increasing number of (agricultural) biogas production plants. Although biogas is currently mainly used in combination with heat and power stations (CHP; production of electricity at the biogas production site), biogas is increasingly used as vehicle fuel and injected into local and national gas grids (Weiland 2010). Hence, there is an upcoming market in which biogas is combusted alone or as part of biogenic/fossil (in short: biofossil) fuel mixtures. Verification of the biogenic carbon composition of produced biomethane, biofossil gas mixtures, and of related CO₂ emissions, using preferably the ¹⁴C method, becomes more relevant.

This article will discuss to what extent two different aspects within the ¹⁴C method introduce (systematic) errors in the calculated bioC fraction when applied to fuel mixtures containing biogas and

^{1.} Centre for Isotope Research, ESRIG, University of Groningen, Nijenborgh 4, 9747 AG Groningen, the Netherlands.

^{2.} Corresponding author. Email: s.w.l.palstra@rug.nl.

natural gas. One of the investigated aspects is the uncertainty in the 14C reference value for 100% biogas carbon (its methane fraction in particular). To calculate the bioC fraction of a sample, the ¹⁴C value of this sample has to be compared with the ¹⁴C value of the bioC fraction of the sample (reference value). The ¹⁴C value of biogenic carbon is in general determined by the ¹⁴C value of the atmospheric CO₂ during the time period of photosynthesis by plant material. The ¹⁴C values of the atmosphere for the period 1950 until present have shown large temporal and spatial variations (Levin et al. 1995; Meijer et al. 1995; Hua et al. 2013). Therefore, the 14C reference value used to calculate the bioC fraction varies considerably between different biogenic materials (Fellner and Rechberger 2009; Mohn et al. 2012). Especially when biogas itself needs to be verified for its 100% bioC content or when biogas and fossil gas are already mixed, the 14C reference value of the bioC fraction in these kinds of samples cannot be determined based on direct 14C measurement of the sample itself, but needs to be approximated based on (most likely) atmospheric ¹⁴CO₂ values. This approximation introduces an uncertainty in the calculated bioC fraction, which increases when less information is available about the composition and origin (average harvest year) of the bioC material in the investigated sample. This is shown in this study for eight different biogas samples from different production sites. In the applied approach, the investigated biogas samples have been divided into four different biotype groups with a certain approximated ¹⁴C reference value and a certain uncertainty for each, merely based on the expected harvest year of the biogenic materials used in the specific biogas production processes.

Another aspect that has been investigated in this study, and which has not been investigated so far for any kind of biofossil fuel mixture, is isotope fractionation correction of measured 14 C values. This aspect within the 14 C method can cause systematic errors in the calculated bioC fraction if not applied well. To compare the measured 14 C value of a sample with a 14 C reference value for 100% bioC, both 14 C values should be calculated according to the same specific standardized calculations. 13 C-based isotope fractionation correction (to a normalized value) should be part of these calculations, especially if the biogenic carbon sources of the measured sample and the 14 C reference value are different and have different δ^{13} C values. Norton and Devlin (2006) have shown examples of deviations in the calculated bioC fraction if the isotope fractionation correction has only been applied to the 14 C reference value but not to the measured 14 C value of the sample itself.

When applied, the isotope fractionation correction should in principle only correct for isotope fractionation that has occurred in the ¹⁴C-containing carbon source. Hence, for ¹⁴C applications in which mixtures of fossil and biogenic carbon are analyzed, the isotope fractionation correction should only correct for isotope fractionation that has occurred in the bioC fraction (until measurement). It is, however, often not possible to investigate the δ^{13} C values of the bioC and fossilC fractions separately. In general, only the δ^{13} C value of the final mixture will be analyzed, including the contribution of the fossil carbon. Mook (1980) has described this ¹³C "mixing effect" on the isotope fractionation correction for atmospheric CO₂ samples. de Rooij et al. (2008) present a calculation study for (modern) carbon accelerator mass spectrometry (AMS) samples diluted with fossil carbon before graphitization. The ¹³C mixing effect on the isotope fractionation correction has not been investigated yet for different fuels and flue gas CO2 samples. This effect will in general not be relevant in case the biogenic carbon and fossil carbon have very similar δ^{13} C values (like wood and coal) and when the fraction of fossil carbon is relatively small. Natural gas and biogas (their methane fractions), however, both show a large variation in δ^{13} C values, with values ranging between -25%and -80% (Levin et al. 1993; Bergamaschi et al. 1998; Milkov 2005; Laukenmann et al. 2010). Gas fuel mixtures containing biogas and/or natural gas might therefore show significant ¹³C mixing effects on the isotope fractionation correction, as the measured δ^{13} C value of the biofossil mixture can be very different from the δ^{13} C value of the bioC fraction in the sample. Hence, for these kinds of mixtures some systematic deviation in the ¹⁴C-based bioC fraction can be expected. This study quantifies these deviations, both theoretically and experimentally, for different biogas and natural gas fuel mixtures. The necessary calculations will be presented in detail and the results are tested and illustrated with actual biogas/natural gas mixtures. Finally, this article will show how both investigated aspects together affect the accuracy of the ¹⁴C method for biogas/natural gas mixtures.

EXPERIMENTAL METHODS

To determine the 14 C values of biogas from different production plants and to investigate the isotope fractionation correction of biofossil gas mixtures, different biogas and natural gas samples have been acquired, pretreated, and then combusted to pure CO_2 . The δ^{13} C value of this CO_2 has been analyzed by isotope ratio mass spectrometry (IRMS) and, after graphitization, its 14 C value by AMS. To investigate the isotope fractionation correction, two different mixture series of biogenic and fossil CO_2 (both from combusted CH_4 samples) have been made: one from biogenic and fossil CO_2 with very different δ^{13} C values and one from biogenic and fossil CO_2 with similar δ^{13} C values. For every pair of biogenic and fossil CO_2 , at least five different CO_2 mixtures with bioC fractions between 0 and 100% were made and analyzed on δ^{13} C and 14 C.

Biogas and Natural Gas Samples

Eight raw biogas samples and three natural gas samples have been obtained in 2011 from DNV KEMA, Groningen, the Netherlands. DNV KEMA sampled the gases at production sites (except "Groningen gas," which was sampled in the laboratory of KEMA itself) and determined the composition of the gases based on gas chromatography (GC) analysis in their laboratory. The gas was sampled with 3-L gasbags of various types (SKC Tedlar bags with polypropylene fitting, SKC Flex Foil bags with polypropylene fitting, and Tedlar bags from BaSystemen with polypropylene fitting).

Gas Sample Pretreatment

Before the gas samples were combusted to CO₂, all samples have been pretreated. Part of the pretreatment of the biogas samples was different from that of the natural gas samples, due to the larger CO₂ fraction present in the (raw) biogas samples compared to the natural gas samples. The investigated raw biogas samples contained 8–42% CO₂ (mol. fraction), while the natural gas samples contained maximal 1–3% CO₂. In this study, the CO₂ fraction in the biogas samples has been separated from the CH₄ fraction (and some other minor components), to gain insight into the isotopic carbon composition of both carbon components separately. As the removal of the CO₂ fraction is common practice in the pretreatment of biogas to obtain a safe, energy-efficient, and reliable energy source (biomethane or what is often called "green gas"), the isotope measurement results of the combusted CH₄ fraction are also a realistic example of carbon isotope results that can be expected if pretreated biogas samples are combusted.

In this study, 360–490 mL of raw biogas was prepared, delivering 250–300 mL CO₂ after combustion of the CH₄ fraction. This amount of CO₂ was necessary because it formed the basis of our investigated biogas-natural gas mixtures. To separate the CO₂ and CH₄ fractions of a raw biogas sample, the sample was introduced in a vacuum-pumped system, water and heavy molecules were cryogenically trapped with dry ice/ethanol (–78°C), and the CO₂ was cryogenically trapped in an iso-pentane trap (–160°C). The remaining CH₄ fraction was cryogenically trapped with liquid N₂ (–196°C) in a 20-mL flask that contained approximately 4 g of molecular sieve 5A. The trapped CO₂ fraction was transferred to a cylinder. To obtain sufficient CH₄ for the combustion process, the CO₂-CH₄ separation procedure was performed twice due to size limitations of the system. The absorbed CH₄ fraction in the 20-mL flask was desorbed from the molecular sieve material (by warming it up)

while connected to a vacuum-pumped 2.5-L flask. The obtained gas sample in the 2.5-L flask was diluted with pure N_2 gas to 1 atm. to make it suitable for the combustion system and to obtain a low methane concentration (to enhance the combustion efficiency in the specific combustion system at a certain gas flow rate). The CH_4 fraction of the biogas sample was then ready to be combusted.

For natural gas samples, the procedure prior to combustion was slightly different, as the separation of CO₂ was not necessary. Only water (if present) and heavy molecules have been removed from the natural gas samples by trapping them cryogenically in a vacuum-pumped system with a dry ice/ethanol mixture. Approximately 380 mL of gas was transferred to a 2.5-L flask and, like the biogas samples, this flask was filled to 1 atm. with pure nitrogen gas as well.

Gas Sample Combustion

The combustion method and the procedure to obtain pure CO_2 was the same for biogas and natural gas samples. After pretreatment, the 2.5-L flask with gas sample was connected in series with a (pure) nitrogen gas flow (~35 mL/min). This flow passed through the flask and then, together with the gas sample, through the entire combustion system. The gas flow (of gas sample + N_2) first passed a volume that was oven-heated to 1000°C. At the end of this heated volume, an O_2 flow was introduced to oxidize the gas sample components. The gas flow then passed a volume filled with CuO (oven-heated to 850°C) to oxidize any formed CO to CO_2 . All formed CO_2 was then cryogenically (liquid N_2) trapped and the remaining gas (mainly N_2 and O_2) was released at the end of the system. For each sample, the combustion process lasted 2 hr because (in general) the obtained CO_2 yield was sufficient (>250 mL). To obtain pure CO_2 for the measurements, the combusted gas flow first passed different steps to remove water and contaminants like oxidized nitrogen and sulfur components, using heated pure Ag (450°C), (acidified) KMn O_4 solution, and cryogen (dry ice/ethanol) water traps. Remaining NO_x components in the obtained CO_2 fraction were removed using pure Cu (at 600°C). All obtained CO_2 samples were stored in vacuum-pumped 200- or 500-mL cylinders.

13C and 14C Analyses of CO₂ Samples

For the isotope measurements, a small aliquot (\sim 5 mL) of each obtained CO₂ sample from the different biogas and natural gas samples was used. This CO₂ volume was cryogenically trapped in a 20-mL flask. For the ¹⁴C measurements of all biogas CO₂ samples, the CO₂ was trapped in 20-mL flasks containing Sulfix (WAKO, 8 \sim 20 mesh). Sulfix removes sulfur-containing compounds by heating the flasks in special flask ovens (at 200°C) during one night. Without this pretreatment, the graphitization of these CO₂ samples was not possible.

 δ^{13} C analyses of all CO₂ samples have been performed with either a SIRA-10 or OPTIMA IRMS. For the 14 C analysis with the AMS system, all CO₂ samples were graphitized to approximately 2 mg graphite and pressed into aluminum targets (Aerts-Bijma et al. 1997). The targets have been analyzed on 12,13,14 C with the 14 C-dedicated AMS (High Voltage Europa Tandetron) of the Centre for Isotope Research laboratory in Groningen (Wijma et al. 1996; van der Plicht et al. 2000).

Biofossil CO₂ Mixtures

Two different series of biofossil CO_2 mixtures have been made, with samples ranging between 0 and 100% bioC. In one series, the $\delta^{13}C$ values of the mixed bioC and fossilC were very different. In the other series, the $\delta^{13}C$ values of both fractions were similar. To make a biofossil CO_2 mixture, CO_2 samples from both gases were expanded into calibrated volumes, each at specific pressure. Both cryogenically trapped portions were subsequently expanded and mixed in the evacuated system and, together, cryogenically trapped and expanded once more to obtain a homogeneous mixture.

The obtained biofossil CO_2 mixture was expanded into two 20-mL flasks. One flask contained Sulfix (see above) and was used for the ^{14}C analysis of the mixture, while the other flask was used for the IRMS $\delta^{13}C$ measurement.

CALCULATION METHODS

Calculation of ¹³C and ¹⁴C Values

The δ^{13} C value of each investigated sample, $^{13}\delta_{sample}$, has been obtained by measuring the 13 C/ 12 C ratio of the sample, relative to the measured 13 C/ 12 C ratio of a local reference material:

$${}^{13}\delta_{sample} = \frac{\binom{13}{12} \binom{12}{12} \binom{13}{12} \binom{12}{12} \binom{13}{12} \binom{$$

The δ^{13} C value of our local reference material with respect to the original VPDB calibration standard (Gonfiantini 1984) is known. All δ^{13} C values shown in this study are expressed with respect to VPDB. The ¹⁴C value of each investigated sample in this study, symbolized here as ¹⁴ a_N^S (in percent, often called pMC), has been calculated as following:

$${}^{14}a_{N}^{S}(pMC) = \frac{{}^{14}A_{N}^{S}}{{}^{14}A_{RN}^{0}} = \frac{\left({}^{14}A_{sample} - {}^{14}A_{bg_{sample}}\right) \cdot \left[\frac{1 + {}^{13}\delta_{N}}{1 + {}^{13}\delta_{sample}}\right]^{2}}{0.7459 \cdot \left({}^{14}A_{OX2} - {}^{14}A_{bg_{OX2}}\right) \cdot \left[\frac{1 + {}^{13}\delta_{N}}{1 + {}^{13}\delta_{OX2}}\right]^{2}} \cdot e^{\lambda \cdot (t_{0} - t_{s})} \cdot 100\%$$

This is the internationally agreed (and often confusing) standardized calculation for the (relative) amount of ¹⁴C in a sample that is used in atmospheric research and other studies. This specific calculation should be used as well in the application to determine the bioC fraction in fuels and other materials. In most published papers about this specific ¹⁴C application, it is often not clear how the reported ¹⁴C values have been calculated exactly. Authors often refer to the paper of Stuiver and Polach (1977), but based on this reference it is still not clear which calculation method has been used exactly. The calculation is different for different ¹⁴C applications (see also Mook and van der Plicht 1999). Results can for instance be reported as pMC values, but might not have been calculated exactly according to Equation 2. LSC ¹⁴C results are often not corrected for isotope fractionation. It is important to point out for those who use the ¹⁴C method to determine bioC fractions, that Equation 2 should always be applied irrespective of the ¹⁴C measurement technique employed. The purpose of this equation is to obtain equal ¹⁴C values for samples with equal biogenic carbon fractions, by correcting for all effects that change the ¹⁴C abundance other than decay between the origin of the substance and the sampling date. Such effects include various isotope fractionation phases and the time between sampling and measurement.

For bioC determination, ${}^{14}a_N^S$ is usually called percentage modern carbon (pMC). The standardization and normalization procedures to obtain these pMC values were first described by Stuiver and Polach (1977). Mook and van der Plicht (1999) updated this description, mainly in the use of symbols and in showing the calculation procedures for different applications. This study shows the symbols as used in Mook and van der Plicht (1999). The ${}^{14}a_N^S$ value symbolizes the measured 14 C signal in the sample relative to the measured 14 C signal in calibration material (a), corrected for isotope fractionation (a) and for decay of the calibration material (a).

In Equation 2,¹⁴ A_N^S is the measured "signal" (abundance or activity depending on the used measurement technique) of ¹⁴C in a certain sample, ¹⁴ A_{sample} . In this study, the ¹⁴C abundance has been determined by calculating the ¹⁴C/¹²C isotope ratio from AMS measurements. ¹⁴ A_{sample} is corrected for the measured ¹⁴C abundance in a specific background sample, ¹⁴ $A_{bg_{sample}}$ (similar pretreatment and measuring method as sample), for isotope fractionation $(1+^{13}\delta_N/1+^{13}\delta_{sample})^2$ and for decay of the calibration material between 1950 (t_0) and the time of harvest of the biogenic sample material (t_s). $\lambda = \ln 2/5730 \text{ yr} = 1/8267 \text{ yr}^{-1}$ is the decay rate of ¹⁴C. ¹³ δ_N is a standardized ¹³ δ C value of -0.025 or -25% (relative to VPDB). ¹³ δ_{sample} is the δ ¹³C value of the sample calculated according to Equation 1.

 $^{14}A_{NN}^{0}$ is the measured 14 C signal (14 C/ 12 C ratio in this study) of the standardized calibration material (oxalic acid II; SRM 4990c). This value has been corrected for the measured background 14 C abundance ($^{14}A_{bgOX_2}$; similar pretreatment and measuring method as the reference standard) for isotope fractionation ($1+^{13}\delta_N/1+^{13}\delta_{OX_2}$)², and corrected with a factor of 0.7459. The latter value is a specific correction factor for the use of the oxalic acid II standard, which is related to other reference materials that were used prior to this standard (Stuiver 1983). $^{13}\delta_{OX_2}$ is the standardized isotope fractionation value of the oxalic acid II standard: -0.0178 or -17.8% (relative to VPDB; Mann 1983). Finally, it has been assumed that both the sample and reference standard have been measured under the same measurement (efficiency) conditions.

Calculation of the bioC Fraction

The bioC fraction (in %) of a sample is calculated as follows:

$$f_{bioC} = \frac{{}^{14}a_{N(sample)}^{S}}{{}^{14}a_{N(100\%bioC)}^{S}} (\times 100; \%)$$
 (3)

where ${}^{14}a_{N(sample)}^{S}$ is the measured 14 C value of the sample and ${}^{14}a_{N(100\%\ bioC)}^{S}$ is the measured or estimated 14 C value of the biogenic carbon fraction in the sample.

Isotope Fractionation Correction

The term $(1^{+13}\delta_N/1^{+13}\delta_{sample})^{\phi}$ in Equation 2 corrects the measured 14 C amount for isotope fractionation based on 13 C isotope ratio measurements to a normalized value, $^{13}\delta_N$ of -0.025 or -25% (relative to VPDB), where, by convention, $\phi = 2$ (although $\phi \approx 2$ as explained in Mook and van der Plicht 1999).

In the normal, routine isotope fractionation correction of an AMS 14 C sample, the δ^{13} C value of the sample ($^{13}\delta_{sample}$) is used as measured with the AMS ($^{13}\delta_{sample} = ^{13}\delta_{AMS}$). In this routine correction, $^{13}\delta_{AMS}$ is measured in the total carbon fraction of the sample. It includes the δ^{13} C contributions of the different carbon components in the sample and the different fractionation effects ($^{13}\alpha$) during the sampling, pretreatment, and measurement processes of the whole sample. In the case of biofossil carbon mixtures, the measured $^{13}\delta_{AMS}$ value thus also includes the fractionation contribution (with $^{13}\alpha = ^{13}\delta + 1$) to the 14 C-free fossil carbon fraction:

$$^{13}\delta_{AMS} + 1 = \left(\left(1 + ^{13}\delta_{bioC} \right) \cdot f_{bioC} + \underbrace{\left(1 + ^{13}\delta_{fosC} \right) \cdot f_{fosC}} \right) \cdot ^{13}\alpha_{sampling} \cdot ^{13}\alpha_{pretreatment} \cdot ^{13}\alpha_{AMS_measurement}$$
(4)

For a correct isotope fractionation correction, the fractionation in the ¹³C signal should be representative for the fractionation in the ¹⁴C signal. This is only the case if the abundances of both ¹³C and ¹⁴C have changed (in a constant ratio) during the same chemical and physical processes. In the

case of biofossil mixtures, the 14 C-free fossil carbon has a certain δ^{13} C value before mixing with the 14 C-containing biogenic carbon. This fossil δ^{13} C value is not related to any fractionation in the 14 C of the mixed biofossil sample. Hence, the 13 C atoms of the fossil carbon fraction and the 14 C atoms of the biogenic fraction have not followed the same fractionation pathways before the mixing of both fractions. The fractionation factor of the fossilC fraction in a sample should therefore be excluded in the fractionation correction. The sample should, in principle, only be corrected based on the δ^{13} C value that represents the fractionation in the bioC fraction of the sample: $^{13}\delta_{AMS\ bioC}$

$$^{13}\delta_{AMS_bioC} = \left(\left(1 + ^{13}\delta_{bioC} \right) \cdot ^{13}\alpha_{sampling_bioC} \cdot ^{13}\alpha_{pretreatment_bioC} \cdot ^{13}\alpha_{AMS_measurement} \right) - 1 \tag{5}$$

If the δ^{13} C values of the bioC and fossilC fractions are not the same, then the use of the routinely measured $^{13}\delta_{AMS}$ value instead of $^{13}\delta_{AMS_bioC}$ will introduce systematic deviations in the calculation of $^{14}a_{N(sample)}^S$ and f_{bioC} . These deviations are likely to occur in mixtures of biogas and natural gas because these gases show, contrary to many solid and liquid fuels (like wood, coal, ethanol), a wide range of δ^{13} C values (as explained in the Introduction).

The best way to avoid these systematic deviations would be to correct with $^{13}\delta_{AMS_bioC}$ instead of $^{13}\delta_{AMS}$. However, $^{13}\delta_{AMS_bioC}$ cannot be measured separately from $^{13}\delta_{AMS}$ in already mixed biofossil samples. Approximation of this value based on the 13 C measurement of the sample's bioC fraction is for most mixtures not possible either. The bioC fraction of biofossil mixtures is usually not separately available for measurement on its carbon isotope composition. The use of $^{13}\delta_{AMS}$ and the related introduction of a deviation in f_{bioC} are therefore inevitable for biofossil gas mixtures.

Calculation of the Deviation in the bioC Fraction

The absolute deviation in the bioC fraction due to the isotope fractionation correction (using $^{13}\delta_{AMS}$ instead of $^{13}\delta_{AMS\ bioC}$) was calculated as follows:

abs. deviation in
$$f_{bioC(^{13}\delta_{AMS})} = f_{bioC(^{13}\delta_{AMS})} - f_{bioC(^{13}\delta_{AMS_bioC})}$$
 (×100; %) (6)

with

$$f_{bioC(^{13}\delta_{AMS})} = \frac{^{14}a_{(sample)}^{S}}{^{14}a_{N(100\%\,bioC)}^{S}} \cdot \left[\frac{0.975}{(1 + ^{13}\delta_{AMS})} \right]^{2}$$
 (7)

and

$$f_{bioC(^{13}\delta_{AMS_bioC})} = \frac{^{14}a_{(sample)}^{S}}{^{14}a_{N(100\%bioC)}^{S}} \cdot \left[\frac{0.975}{\left(1 + {^{13}\delta_{AMS_bioC}}\right)} \right]^{2}$$
(8)

If no fractionation correction is applied at all, the absolute deviation in the bioC fraction is

abs. deviation in
$$f_{bioC(nocorr)} = f_{bioC(nocorr)} - f_{bioC(^{13}\delta_{AMS,bioC})}$$
 (9)

with

$$f_{bioC(nocorr)} = \frac{{}^{14}a_{(sample)}^{S}}{{}^{14}a_{N(100\%bioC)}^{S}}$$
(10)

This study calculates the deviations based on real 13 C and 14 C measurements of biofossil mixtures and based on different theoretical cases. When calculating the theoretical cases, the δ^{13} C value of each theoretical sample (with a certain bioC fraction between 0 and 100%) has been determined based on the chosen δ^{13} C values of the 100% bioC and 100% fossilC fractions and the chosen bioC fraction of the sample. For these cases, $^{13}\delta_{AMS}$ is replaced by $^{13}\delta_{biofossil\,sample}$ and $^{13}\delta_{AMS_bioC}$ by $^{13}\delta_{100\%\,bioC}$ in Equations 6–9. To calculate $f_{bioC}(^{13}\delta_{AMS_bioC})$ (Equation 8) for the real measurements, $^{13}\delta_{AMS_bioC}$ (Equation 5) was approximated as follows, because it could not be measured directly in the mixed biofossil samples:

$${}^{13}\delta_{AMS_bioC} \cong \left(\left(1 + {}^{13}\delta_{IRMS_100\%bioC} \right) \cdot \frac{\left(1 + {}^{13}\delta_{AMS} \right)}{\left(1 + {}^{13}\delta_{IRMS} \right)} \right) - 1 \tag{11}$$

with

$$^{13}\delta_{IRMS_100\%bioC} = \left(\left(1 + ^{13}\delta_{bioC} \right) \cdot ^{13}\alpha_{sampling_bioC} \cdot ^{13}\alpha_{pretreatment_bioC} \right) \tag{12}$$

 $^{13}\delta_{IRMS_100\%bioC}$ is the δ^{13} C value of the separately available and measured bioC material (that was used to make a biofossil mixture). It represents the δ^{13} C value of the bioC material in the mixed biofossil sample after sampling, pretreatment, and combustion to CO₂, and

$$\frac{\left(1+\frac{13}{\delta_{AMS}}\right)}{\left(1+\frac{13}{\delta_{IRMS}}\right)} = {}^{13}\alpha_{AMS_measurement}$$
(13)

where $^{13}\alpha_{AMS_measurement}$ represents the fractionation in each measured biofossil mixture after graphitization (which is minimal according to Aerts-Bijma et al. 1997) and AMS measurement. $^{13}\delta_{IRMS}$ is the δ^{13} C value of the biofossil sample before it is graphitized and $^{13}\delta_{AMS}$ is the δ^{13} C value of the sample after graphitization and measurement with AMS.

RESULTS AND DISCUSSION

This section first gives an overview of the origin and composition of the different biogas and natural gas samples that have been investigated in this study. The ¹³C (IRMS) and ¹⁴C (AMS) measurement results are given for the separated CO₂ and (combusted) CH₄ fractions of the eight biogas samples and for the measured CO₂ of the three combusted natural gas samples. The measured carbon isotope values are discussed in detail and the results compared with other studies. A new approach is then presented to determine the ¹⁴C reference values for different biogas samples. The implications of this approach are considered for the accuracy of the bioC fraction calculation. Next, systematic deviations in the calculated bioC fraction due to departures from the correct isotope fractionation correction are presented and discussed. Finally, examples are given of possible errors in the calculated bioC fraction due to the combination of different error sources.

Origin and Composition of the Investigated Gases

The molecular compositions of the investigated gas samples, as analyzed after sampling and prior to the gas pretreatment procedure, are shown in Table 1. The sample names give information about

the origin or production source of the gases. All biogas samples come from Dutch production plants. The biogas samples Landfill_1 and Landfill_2 are from two different municipal solid waste landfill sites. "Organic waste" is biogas from the digestion of the (wet) organic fraction of municipal waste. "Cookies_fish" is biogas from the waste materials of two different food factories where cookies and fish products are produced. "Mun. sewage sludge" is biogas from municipal sewage sludge. "Manure_vegetables" has been produced from manure together with grass, maize, vegetable fat, and fried potatoes. The biogases Sugar beet and Manure_vegetables are used in combined heat and power (CHP) stations. The other biogases are injected into Dutch national distribution gas networks after pretreatment. The exact production sites of the natural gas samples are classified (by DNV KEMA). Norway gas is a high-caloric gas (caloric value > 37 MJ/m³); all other bio- and natural gases have are low-caloric gases.

¹³C and ¹⁴C Measurement Results Biogas and Natural Gas

The results of the 13 C and 14 C analyses for the different biogas and natural gas samples are listed in Table 2. The δ^{13} C (with respect to VPDB) and $^{14}a_N^S$ values have been calculated according to Equations 1 and 2, respectively.

The CO₂ fraction of the Mun. sewage sludge has not been analyzed by ¹⁴C as the (cryogenic) extraction yield of CO₂ from this biogas was too low for ¹⁴C analysis. The yield was lower than expected based on the composition of the biogas. This was also the case for the CO₂ yield of the combusted CH₄ fraction of this biogas sample. A clear reason for this has not been found. The combustion of this gas contaminated the combustion system severely. Contamination might also have played a role in the problematic extraction of the CO₂ fraction of this biogas sample (this has however not been investigated further).

The absolute measurement errors (1 σ) are $\pm 0.03\%$ for all δ^{13} C values (IRMS), $\pm 0.3-0.5\%$ for the $^{14}a_N^S$ values of the biogas samples, and $\pm 0.10\%$ for the $^{14}a_N^S$ values of the natural gas samples. The reproducibility of the applied combustion method was checked by performing multiple combustions for two of the natural gases (Norway gas and Groningen gas). All individual results (for both 13 C

Table 1 Main molecular composition (in mol%) of the investigated biogas samples and natural gas samples prior to pretreatment in the laboratory (measurements by DNV KEMA).

Biogas	CH ₄ (mol%)	CO ₂ (mol%)	C_xH_y (mol%)	(N ₂ , O ₂ ,) (mol%)
Maize_onions	56.27	42.20	0.02	1.52
Landfill_1	61.13	32.12	0.00	6.75
Organic waste	61.47	38.35	0.00	0.18
Cookies_fish	85.24	11.96	0.00	2.81
Mun. sewage sludge	89.49	10.25	0.00	0.26
Landfill_2	54.43	30.47	0.00	15.10
Sugar Beet	87.75	8.15	0.00	4.07
Manure_vegetables	59.56	35.75	0.00	4.70
Natural gas				
Norway gas	87.10	2.06	8.90	1.95
North Sea gas	85.07	2.66	4.93	7.34
Groningen gas	81.05	1.02	3.45	14.48

and 14 C) of these multiple series correspond within 2 times the given measurement error. The (average) $^{14}a_N^S$ value of the Norway gas is relatively high for a fossil gas that contains in principle no 14 C. Although the Norway gas value is still at background level, the possibility of contamination of the samples prior (mixed with biogas?) or during sampling cannot be excluded.

δ13C Values

The δ^{13} C values of the combusted CH₄ fractions ("CO₂ after combustion" in Table 2) of seven different biogas samples vary between -39.2% and -61.3%. This is comparable to the results of studies by Laukenmann et al. (2010) and Levin et al. (2003), who showed δ^{13} C values between -50% and -70% for biogas methane from relatively similar digestion processes. The δ^{13} C value of -28.6% for Mun. sewage sludge is relatively high compared to that of the other investigated biogas samples.

The observed variation between the biogas samples is mainly related to the differences in the anaerobic digestion processes of the biogas production plants. The CH₄ in all investigated biogas samples has been produced by anaerobic digestion of the organic input materials by specific microorganisms (like methanogens; for landfill and sewage sludge also other groups are involved). The main biogas CH₄ formation pathways are the production of methane from acetate by acetotrophic methanogens and from CO₂ reduction by hydrogenotrophic methanogens. Methane obtained from acetate fermentation shows less-depleted δ¹³C values than methane obtained from CO₂ reduction (Whiticar et al. 1986; Krzycki et al. 1987; Conrad 2005). This might explain the less-negative δ^{13} C value of municipal sewage sludge CH₄ (Krzycki et al. 1987). Which pathways occur or dominate the digestion process (and the isotope fractionation) depend on the type and number of microorganisms available in the process and on the parameters that influence these microorganisms and their ability to digest the organic material, like the composition of the organic material and the fermentation temperature (Conrad 2005). Methane from landfill gas has been isotopically analyzed in many different studies (Bergamaschi et al. 1998; Cabral et al. 2010; Zyakun et al. 2010) and its δ^{13} C varies between -30%and -60% and is often < -45%, comparable to the measured values in this study (-61.3% for Landfill 1 and -56.3% for Landfill 2). The variation between different landfill gas samples is, besides the variations in production processes, also due to differences in oxidation rate in time and depth.

Table 2 Determined $^{13}\delta C$ values (IRMS) and $^{14}a_N^S$ values (AMS) of pretreated biogas and natural gas samples after combustion to CO_2 , and of the CO_2 fraction of raw biogas samples.

	CO ₂ after combustion		CO ₂ fraction raw biogas		
Biogas	δ ¹³ C (‰)	$^{14}a_{N}^{S}\left(\mathrm{pMC}\right)$	δ ¹³ C (‰)	$^{14}a_N^S (pMC)$	
Maize_onions	-55.58	102.6	26.57	102.3	
Landfill_1	-61.24	104.0	14.96	104.2	
Organic waste	-52.04	104.5	3.14	104.1	
Cookies_fish	-53.64	104.3	30.45	104.2	
Mun. sewage sludge	-28.55	105.1	11.72	_	
Landfill_2	-56.25	116.1	6.58	115.8	
Sugar beet	-39.20	102.3	-5.55	101.8	
Manure_vegetables	-48.43	103.4	8.05	102.8	
Natural gas					
Norway gas	-39.86	0.18	_	_	
North Sea gas	-29.81	0.10	_	_	
Groningen gas	-28.38	0.05	_	_	

The investigated CO_2 fractions of the biogas samples show enriched $\delta^{13}C$ values that vary between -5.6% for Sugar beet and +30.5% for the Cookies_fish biogas sample. The enrichment of the CO_2 is related to the formation process of the depleted CH_4 (Conrad 2005) and has been measured in other studies as well (Rosenfeld and Silverman 1959; Levin et al. 1993; Laukenmann et al. 2010; Zyakun et al. 2010).

The overall δ^{13} C value of the biogas carbon fraction ($^{13}\delta_{biogas} \cong ^{13}\delta_{CH_4} \cdot f_{CH_4} + ^{13}\delta_{CO_2} \cdot f_{CO_2}$) varies for the investigated biogas samples between -20% (Maize_onions) and -43% (Cookies_fish; perhaps due to different kind of reservoir effects?).

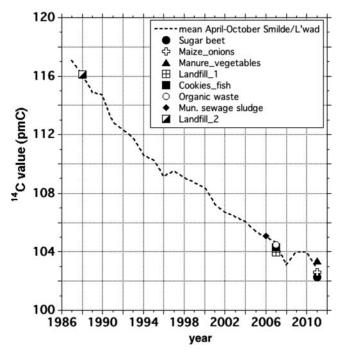
The δ^{13} C values of the investigated natural gases vary between -28% and -40%. The variation is small compared to the large range of δ^{13} C values between -25% and -80% measured in natural gas samples at different sites (Schoell 1980; Milkov 2005). The variation in δ^{13} C values of natural gas samples is due to the different and very variable formation processes of the gases (Fuex 1977). The measured δ^{13} C value of the Groningen gas (-28.4%) is close to the -29.0% found by Hut et al. (1984) for Groningen natural gas sampled at A.F. Tjuchem in the Netherlands.

¹⁴C Values Of Biogas

The 14 C values ($^{14}a_N^S$) of the carbon components (CH₄, CO₂) in biogas are mainly determined by where and when the specific carbon has been taken up as atmospheric CO₂ by plant photosynthesis. Over the last 60 yr, the $^{14}a_N^S$ values of atmospheric CO₂, and thus the $^{14}a_N^S$ values of organic material, have shown large temporal and spatial variations (the maximum value reached in 1964 is ~90 pMC higher than today's atmosphere). Hua et al. (2013) recently made an overview of (average) atmospheric 14 CO₂ values on a global scale in the period 1950–2010. Fellner and Rechberger (2009) have investigated the variation in $^{14}a_N^S$ values of the different components in waste and solid recovered fuel (SRF) (like wood, paper, plastics, vegetables). In their study, the $^{14}a_N^S$ values range between 98 and 135 pMC, depending on the type and mixture of organic materials. The $^{14}a_N^S$ values of seven of the biogas samples investigated in this study (CO₂ and combusted CH₄ fractions) range between 102 and 105 pMC, while one biogas sample (Landfill 2) is considerably higher, at 116 pMC.

Figure 1 shows where the $^{14}a_N^S$ values of the investigated biogas samples (results of the combusted CH₄ fraction) can be found on the timeline of annual average monthly mean atmospheric $^{14}a_N^S$ values (average April-October) measured at the Dutch monitoring stations Smilde (1987–2003) and Lutjewad (2003–2011). It gives an impression of the average year of atmospheric 14 CO₂ sampling by plants, and thus for the carbon of the organic materials that have been used in the production of the investigated biogas. As all investigated biogas samples have been produced in the Netherlands, it is assumed that the products originate from plants that have grown in the Netherlands as well. The (rural) Dutch monitoring sites are therefore assumed to be representative for the sites of plant growth.

For the biogas sample Sugar beet, it was known that it was produced from sugar beets that were grown, harvested, and digested in 2011 in the Netherlands. This is confirmed by the data in Figure 1 in which the $^{14}a_N^S$ value of this biogas sample is the same as the average value measured in Dutch atmospheric air in 2011. For the other biogas samples, it is expected that digested vegetables will in general have $^{14}\text{CO}_2$ sampling (and harvest) years that are close to the year of digestion as they are usually not stored for a long time. This seems to be the case for the Maize_onions sample and also for Manure_vegetables, although the latter also matches the atmospheric 2008 value. Manure carbon could have been from earlier years than the digested vegetables. For municipal sewage sludge and digested food products (like cookies), the average $^{14}\text{CO}_2$ sampling (and harvest) year deviates a few years from the year of digestion (2011). This can be expected because the organic material is



Comparison between 14C values Figure 1 (pMC) of different biogas samples and average atmospheric 14C values (pMC) for the period 1987-2011. The atmospheric ¹⁴CO₂ values have been measured at Dutch monitoring stations Smilde (1987-2002; published before in Meijer et al. 1995 and Palstra et al. 2008) and Lutjewad (2004-2011) by the University of Groningen. The biogas results are from combusted CH4 fractions. The plotted annual atmospheric 14C values are average values from monthly mean 14CO2 values of the months April-October. The period in which plants take up atmospheric CO2 is typically within this period.

not directly digested after harvest, but has first been used in other processes during a certain time period. An example of long-term storage (decades) of atmospheric carbon is wood-based material. In the case of biogas, this long-term storage can be found in organic material stored and "digested" at old landfill sites, as the result of Landfill_2 shows (116 pMC; average atmospheric carbon year is 1988). The storage effect of wood-based plants can also to some extent be seen at landfill sites that are still in use (Landfill_1) and in organic waste, which both contain municipal organic waste fractions that have been recently dumped.

When determining the ¹⁴C values ($^{14}a_{N(sample)}^S$; Equation 2), the year of harvest of the biogenic organic material, t_s , has to be filled in as well. If biogas or biofossil gas mixtures need to be verified and the average harvest year is not known, then the year of digestion should be used instead. Fortunately, the term in Equation 2 containing the harvest year varies very little per year. The calculated $^{14}a_N^S$ value deviates only 0.01% (relatively) with every year of difference between the year of digestion and the average harvest year. For the investigated biogas samples of this study with unknown harvest year, t_s = 2011 was used, because that was the year of digestion. For the Sugar beet samples, it was known that the harvest year was 2011. If the harvest years for the "unknown" samples are estimated based on Figure 1, the use of 2011 instead of the "real" harvest year gives deviations of <0.05 pMC for most of the biogas samples. Only for Landfill 2, for which the harvest year (or "average year of growth") 1988 seems to be a good estimate, $^{14}a_{N(sample)}^S$ would increase distinctively, by 0.3 pMC (from 116.1 to 116.4 pMC) if 1988 had been used for t_s instead of 2011.

The 14 C values of the separately measured CO₂ fraction and combusted CH₄ fraction of biogas samples agree for all investigated biogas samples within two times the measurement error. This is also expected, as both fractions originate from the same digested organic materials. In cases of fraud investigation to identify whether biomethane is 100% biogenic, the measurement of the CO₂ fraction could therefore act as an internal reference (comparison with the $^{14}a_N^S$ value of the combusted CH₄ fraction). The observed differences in $^{14}a_N^S$ values between both carbon fractions in this study (0.2–

 $0.6 \,\mathrm{pMC}$) can be due to other factors than the random measurement errors. For example, variation in the $\mathrm{CO}_2/\mathrm{CH}_4$ ratio (mol %) between digested organic materials with different ${}^{14}a_N^S$ values (within the same biogas batch) give different average ${}^{14}a_N^S$ values of CO_2 and CH_4 in a particular biogas sample.

¹⁴C Reference Value for 100% Biogenic Carbon

When determining the bioC fraction of biogas and biofossil gas mixtures (Equation 3), a ¹⁴C reference value for 100% biogenic carbon is needed ($^{14}a_{N(100\% bioC)}^{S}$). Ideally, this value would be the measured ¹⁴C value of the pure biogenic gas that lies at the basis of the sample. Usually, however, this measurement is not possible or feasible, as in most cases the biofossil carbon samples are already mixed (and/or combusted). For biogas samples that need to be verified for bioC composition, a reference value for 100% bioC should be available beforehand and cannot be determined based on the measurement of the biogas sample itself. Therefore, in most cases an approximated value needs to be used that is based on ¹⁴C values of different biogenic materials or based on atmospheric ¹⁴CO₂. An approximated ${}^{14}a_{N(100\%bioC)}^{S}$ value will, obviously, introduce a bigger uncertainty in the calculated bioC fraction (f_{bioC}) than a directly measured one. Especially for fraud investigation, where fraud should be significantly identified from systematic and random uncertainties in the determined bioC fraction, it is of interest that the approximated value is reliable and that the uncertainty in this value is as small as possible. Quantification of uncertainties in the calculated bioC fraction, including the uncertainty in the used ${}^{14}a_{N(100\%bioC)}^S$ value, is therefore essential. So far, however, it has not been part of the several (inter)-national standards in which the 14C method is used to determine the bioC fraction (like ASTM D6866-12 [ASTM 2012] and ISO/FDIS 13833 [ISO 2012]).

This study proposes a method to approximate $^{14}a_{N(100\%bioC)}^{S}$ values for different types of biogases based on atmospheric $^{14}\text{CO}_2$ values. Biogases were divided into four different groups with different (maximum) time periods between growth/harvest and digestion. For each group, the $^{14}a_{N(100\%bioC)}^{S}$ value is calculated based on atmospheric $^{14}\text{CO}_2$ values that have been measured in the defined time period of that group. The defined groups are summarized in Table 3. The (average) year of plant growth is known only for group 1. For the other groups, this is not known and thus has been estimated based on information about the used organic materials: Group 2 is organic material with atmospheric carbon from the harvest year only, digested within 2 yr after harvest. Group 3 consists of two subgroups of organic materials. The first subgroup is organic material with atmospheric carbon from the harvest year only, but which has been pretreated first before becoming a waste product. The second subgroup is organic material stored in plants for a short average time period (few years). Both subgroups are characterized by an average of up to 5-yr difference in time between harvest and digestion of the material. Group 4 is the most complicated one. It contains mixtures of organic material, stored for a long time period (up to several decades) until digestion to biogas (municipal waste is a typical example).

This method can also be used for other (solid, liquid) biofuels. Instead of the year of digestion, the year of biofuel production can then be used. Table 4 lists 100% bioC reference values [$^{14}a_{N(100\%bioC)}^{S}$] for biogases digested in 2011, which have been calculated according to the approach of Table 3. For groups 1–3, values were calculated based on averaged monthly mean atmospheric $^{14}\text{CO}_2$ values measured at the Dutch monitoring stations Smilde (2001–2003) and Lutjewad (2004–2011) (Centre for Isotope Research, University of Groningen; monthly mean values have not been published yet). For each group, the average ^{14}C value has been calculated based on monthly data from the period April–October for the year(s) in the given time period of the specific group. The uncertainties given are the standard deviations in the averaged monthly mean values. The (rounded off) $^{14}a_{N(100\%bioC)}^{S}$ value and its overall uncertainty for biogas group 4 have been based on (waste) data by Fellner and Rechberger (2009; see their Table 5).

Table 3 Approach to approximate the 100% bioC 14 C reference value ($^{14}a_{N(100\%bioC)}^{S}$) for different biogas groups.

Biogas group	Approach
1. (Average) year of plant growth is known.	Average atmospheric ¹⁴ CO ₂ value April–October of the (average) year of plant growth.
2. Year of plant growth is not known and is expected to differ <2 yr with year of digestion (vegetables, annual grown plants; no wood).	Average atmospheric ¹⁴ CO ₂ value April–October of the year of digestion and the previous year.
3. (Average) year of plant growth is not known and average difference between biomass harvest and digestion is estimated to be <10 yr (food residue, manure, sewage sludge, wet organic municipal waste fraction; wood <10 yr).	Average atmospheric ¹⁴ CO ₂ values April–October of 10 yr incl. year of digestion.
4. (Average) year of plant growth is not known and average difference between biomass harvest and digestion >10 yr (municipal waste stored in old landfills, wood >10 yr).	Average ¹⁴ C value of waste materials in year of sampling, based on study by Fellner and Rechberger (2009).

The values in Table 4 are valid for 2011 only. Due to the (long-term) annually decreasing $^{14}a_N^S$ value of atmospheric CO₂ (since the 1960s), the approximated $^{14}a_{N(100\%bioC)}^S$ values of the different groups will decrease annually as well (only 1-yr average values of successive years can show an increase sometimes, as can be seen in Figure 1). This long-term decrease is currently about -0.5 pMC per year (also for waste according to Fellner and Rechberger 2009). Biogas samples digested in 2012 might therefore have slightly lower $^{14}a_{N(100\%bioC)}^S$ values than those mentioned in Table 4.

As the values of groups 1–3 in Table 4 have been calculated based on atmospheric ¹⁴CO₂ values measured in the Netherlands, they represent 14C values that can be found in organic carbon from plant materials that have grown in the Netherlands. For digested organic materials that have grown in other regions/countries, their ${}^{14}a_{N(100\%bioC)}^{S}$ values can be different compared to the Dutch materials, as the ¹⁴a_N values of atmospheric ¹⁴CO₂ and therefore of plant materials as well show spatial variation (Meijer et al. 1995; Levin et al. 2003; Hsueh et al. 2007; Palstra et al. 2008). The $^{14}a_{N100\%bioC}^{N}$ values based on average ¹⁴a_N values of atmospheric ¹⁴CO₂ could therefore be slightly different as well if based on atmospheric values of other regions. A comparison of the differences between averaged $^{14}a_N^S$ values of the Dutch monitoring stations, the remote Alpine monitoring station Jungfraujoch, and highly urbanized regions (Meijer et al. 1995; Levin et al. 2003; Levin and Kromer 2004; Palstra et al. 2008; data Jungfraujoch >2003 from personal communication with I Levin, IUP Heidelberg), shows, however, that the Dutch data are in between the values from (European) remote and highly urbanized regions (± 1.5 pMC). The approximated $^{14}a_{N(100\% bioC)}^{S}$ values in Table 4 are therefore useful average values for biogas samples, regardless of its organic carbon origin (at least in Europe). Only the uncertainties in Table 4 will be slightly larger for biogas groups 1-3 due to this spatial variation (with 1σ approximately ± 0.5 pMC). The total (1σ) uncertainties in the approximated $^{14}a_{N(100\%bloC)}^{S}$ values of biogas groups 1–3 are then ± 0.7 , ± 0.9 , and ± 1.7 pMC, respectively, instead of the uncertainties given in Table 4.

It was verified that the 100% bioC reference values of Table 4 give the correct bioC fractions for the investigated biogas samples (Table 2), within the given uncertainty ranges. For this verification, it was assumed that all investigated biogas samples contained 100% biogenic carbon. As the investigated biogas samples in this study contain organic carbon from plants materials that have mainly

of sumpled in 2011.				
Biogas group	Time period	$^{14}a_{N(100\%bioC)}^{S}(pMC)$	±(1σ)	
1	2011	102.8	0.5	
2	2010–2011	103.4	0.7	
3	2001–2011	105.0	1.6	
4	Not defined (wood materials >1900 AD)	115	4	

Table 4 Approximated ${}^{14}a_{N(100\%bioC)}^S$ values for (Dutch) biogas samples digested and/or sampled in 2011.

grown in the Netherlands, the values and the given uncertainties in Table 4 are expected to be very representative for the investigated biogas samples. The biogas samples have been divided over the four defined groups based on the available information about the organic material used in the biogas production. Table 5 shows the calculated bioC fractions of the biogas samples. The 1σ error in the calculated bioC fraction is determined by the (AMS) measurement error in $^{14}a_{N(100\%bioC)}^{S}$ (0.4 pMC for all samples) and by the uncertainty in the approximated $^{14}a_{N(100\%bioC)}^{S}$ value.

The approach employed to approximate the $^{14}a_{N(100\%bioC)}^S$ value for different groups of biogases works very well for the biogases investigated in this study. All biogas samples contain 100% bioC within 1σ uncertainty and within $\pm 1\%$. Dividing the different biogas samples into groups with different approximated $^{14}a_{N(100\%bioC)}^S$ values improves the accuracy of the 14 C method for the different types of biogas samples. The more information available about the average harvest year and the location and time period of atmospheric carbon uptake, the smaller the uncertainty and systematic deviation in the determined bioC fraction.

Biogas from vegetables that have grown in greenhouses where fossil CO₂ has been added to the air, forming a special class of "bio material." The ¹⁴C values of these plants can in principle be much lower than atmospheric ¹⁴CO₂ values due to the dilution with ¹⁴C-free fossil carbon. If the bioC fraction of these (pure) biogases is calculated using a 100% bioC reference value, as determined according to the method described in Table 3, this bioC fraction will be lower than 100%. On the one hand, this is not satisfactory since the formed gas is purely "biogenic," that is, recently formed by photosynthesis. On the other hand, the ¹⁴C method correctly shows that fossil fuel CO₂ is involved, and is right in the sense that using this biogas will only partially prevent the emission of fossil fuel CO₂ into the atmosphere. What is desirable in this application of the ¹⁴C method for biogas samples is a matter of definition, but both practical and principal arguments are in favor of the correctness of the ¹⁴C-based determination.

Table 5	BioC fraction	(f bioC) of the	investigated	biogas samples.
I doic 5	Dioc muchom	(1 bloc) of the	mvestigatea	orogus sumpres.

Biogas	Group nr	f bioC (%)	± (1σ)	Deviation with 100% bioC
Sugar beet	1	99.5	0.6	-0.5
Maize_onions	2	99.2	0.8	-0.8
Manure_vegetables	2	100.0	0.8	0.0
Landfill_1	3	99.0	1.6	-1.0
Cookies_fish	3	99.4	1.6	-0.6
Organic waste	3	99.5	1.6	-0.5
Mun. sewage sludge	3	100.1	1.6	0.1
Landfill_2	4	101.0	3.5	1.0

Deviations in the bioC Fraction Related to Isotope Fractionation Correction

This section presents deviations in the bioC fraction due to incorrect isotope fractionation correction. Results are given for two experimental and five theoretical cases in which 14 C measurement results ($^{14}a^S$) have been corrected based on $^{13}\delta_{AMS}$ instead of $^{13}\delta_{AMS_bioC}$ for the experimental cases and based on $^{13}\delta_{biofossil\ sample}$ instead of $^{13}\delta_{100\%bioC}$ for the theoretical cases. The experimental cases are based on real 13 C and 14 C measurements of two different mixture series of biogenic and fossil CO₂ (both from combusted CH₄ samples). Deviations have been calculated according to Equation 6 for the theoretical cases and according to Equation 6 combined with Equation 11 (approximated value for $^{13}\delta_{AMS_bioC}$) for the experimental cases. Another two theoretical cases, in which no isotope fractionation correction has been applied, have been investigated as well (using Equation 9). The $\delta^{13}C$ values of the separate bioC and fossilC fractions and the $^{14}a_{N(100\%bioC)}^S$ values that have been used to investigate theoretical biofossil mixtures are summarized in Table 6. For the two experimental cases, the measured $\delta^{13}C$ and (uncorrected for fractionation) $^{14}a^S$ values of different biofossil samples (with bioC fractions ranging between 0 and 100%) are shown in Tables 7 and 8. The chosen values for theoretical cases 1 and 2 (Table 6), are similar to the values of the experimental cases 1 and 2, respectively.

Figure 2 shows the absolute deviations in the calculated bioC fractions for the different investigated experimental and theoretical cases. Isotope fractionation correction based on the δ^{13} C value of biofossil mixtures, instead of the δ^{13} C value of the bioC fraction alone, results in systematic deviations when calculating the bioC fraction of a sample. These deviations increase with increasing difference between δ^{13} C_{bioC} and δ^{13} C_{fosC} and are maximal for samples with 50% bioC. The calculated bioC fraction decreases due to this deviation if δ^{13} C_{bioC} $< \delta^{13}$ C_{fosC} and increases if δ^{13} C_{bioC} $> \delta^{13}$ C_{fosC}. There is no deviation in the calculated bioC fraction if δ^{13} C_{bioC} $= \delta^{13}$ C_{fosC}, as is also shown by experimental case 2 and theoretical case 2. As the δ^{13} C values of biogas and natural gas can vary considerably, but will in general not differ more than 50% (theoretical case 5), absolute deviations in a calculated bioC fraction between 0 and $\pm 2.5\%$ can be expected for biofossil gas mixtures with unknown δ^{13} C values of the bioC and fossilC fractions.

If no isotope fractionation correction is applied (published LSC measurements are often not corrected for isotope fractionation and application of the correction is often questioned by users of LSC-based 14 C data), the systematic deviation in the calculated bioC fraction increases with increasing difference between the δ^{13} C value of the investigated sample and δ^{13} C_N (the normalized δ^{13} C value of -25%). The absolute deviation increases with increasing bioC fraction, is maximal at 100% bioC and can be up to -10% for biogases. Especially if biogas samples are verified for their bioC content (verification of 100% bioC), no fractionation correction of the measured 14 C value in the sample will give erroneous verification results.

Accuracy of the ¹⁴C Method for Biogas/Natural Gas Fuel Mixtures

In the previous sections, two principle sources of error in the calculated bioC fraction have been demonstrated for biofossil gas mixtures: the uncertainty in the 14 C reference value for 100% bioC ($^{14}a_{N(100\%bioC)}^{S}$) and deviations due to isotope fractionation correction. These error sources are part of the 14 C method to determine the bioC fraction, irrespective of the 14 C measurement technique used or the investigated biofossil carbon material. The quantity of the possible errors, however, depends on these latter factors as well. Results are presented that have been measured with AMS, which has, compared to LSC 14 C measurements (Norton and Devlin 2006), a relatively low measurement uncertainty. Reported uncertainties in calculated bioC fractions based on AMS 14 C measurements are therefore more accurate and are not representative for the results based on LSC measurements. The

	$\delta^{13}C_{100\%,fossilC}$ (%0)	$\delta^{13}C_{100\%bioC}$ (%0)	$^{14}a_{N(100\%bioC)}^{S}(pMC)$
Theor. & no corr. case 1	-28.38	-61.24	104.0
Theor. & no corr. case 2	-39.86	-39.20	102.3
Theor. case 3	-50.00	-30.00	100.0
Theor. case 4	-30.00	-40.00	100.0
Theor. case 5	-25.00	-75.00	100.0

Table 6 δ^{13} C values of bioC and fossilC fractions and $^{14}a_{N(100\%bioC)}^{S}$ values, used to calculate deviations in different bioC fractions of theoretical biofossil carbon mixtures.

accuracy of the ¹⁴C method varies between the different ¹⁴C measurement techniques. This should be taken into account in the use of this application for verification purposes.

Figure 3 gives an overview (theoretical cases) of possible errors in the calculated bioC fraction for two groups of biofossil gas mixtures, if measured with AMS, using an approximated reference value for 100% bioC and with unknown δ^{13} C values of the separate biofossil fractions. In Table 3, the different biogas groups are defined. Each group has a certain uncertainty in the 100% bioC reference value ($^{14}a_{N(100\%bioC)}^{S}$). Group 1, biogases with known harvest year, has the lowest uncertainty and group 4, landfill biogas, has the highest uncertainty. In Figure 3, the results of both groups together show the range of possible errors in the 14 C-based method to calculate bioC fractions for biofossil gas mixtures. The possible errors given for groups 1 and 4 are a combination of the (1σ) uncertainties in the 14 C measurement and in the approximated $^{14}a_{N(100\%bioC)}^{S}$ value and the uncertainty in the size of the deviation due to possible incorrect isotope fractionation.

The case "measurement uncertainty" ("meas. uncer." in the graph) shows the 1σ range of the random variation in the calculated bioC fraction due to the AMS measurement uncertainty alone. The values ${}^{14}a_{N(100\%bioC)}^S$ and ${}^{14}a_{N(100\%bioC)}^S$ in the calculation of the bioC fraction (Equation 3) are in this case both directly measured with AMS. The error increases with increasing bioC fraction and reaches

Table 7 Measured δ^{13} C and $^{14}a^S$ values of experimental case 1: CO₂ mixtures of combusted Landfill_1 biogas (δ^{13} C_{IRMS} = -61.24% and $^{14}a^S_{N(100\%bioC)}$ = 104.0 pMC) and combusted Groningen natural gas (δ^{13} C_{IRMS} = -28.38%). Samples 1 and 11 are not mixtures but show the values for the 100% fossilC sample and 100% bioC sample, respectively.

	1				
	¹⁴ a ^S (%)	±	$\delta^{13}C_{IRMS}$ (%0)	$\delta^{13}C_{AMS}$ (‰)	
1 (0% bioC)	0.05	0.04	-28.38	-28.8	
2	2.80	0.05	-29.19	-28.0	
3	5.50	0.07	-30.12	-31.7	
4	6.40	0.08	-30.42	-28.2	
5	22.7	0.1	-36.57	-36.4	
6	45.8	0.2	-43.98	-45.6	
7	64.5	0.3	-49.94	-50.2	
8	70.1	0.3	-52.59	-50.1	
9	90.7	0.4	-59.28	-59.2	
10	95.6	0.4	-61.03	-60.0	
11 (100% bioC)	96.1	0.5	-61.24	-62.6	

Table 8 Measured δ^{13} C and ${}^{14}a^{S}$ values of experimental case 2: CO₂ mixtures of combusted Sugar_beet biogas (δ^{13} C_{IRMS} = -39.20‰) and ${}^{14}a^{S}_{N(100\%bioC)}$ = 102.3 pMC) and combusted Norway natural gas (δ^{13} C_{IRMS} = -39.86‰). Samples 1 and 12 are not mixtures, but show the values for the 100% fossilC sample and 100% bioC sample, respectively.

	$^{14}a^{S}(\%)$	±	$\delta^{13}C_{IRMS}$ (‰)	$\delta^{13}C_{AMS}$ (‰)
1 (0% bioC)	0.16	0.06	-39.86	-40.1
2	1.72	0.05	-39.80	-39.4
3	6.40	0.08	-39.51	-40.7
4	28.5	0.2	-39.62	-41.0
5	31.2	0.2	-39.63	-37.6
6	32.1	0.2	-39.60	-39.1
7	46.5	0.2	-39.48	-40.8
8	69.3	0.3	-39.41	-39.6
9	91.6	0.4	-39.42	-38.7
10	93.5	0.4	-39.26	-38.7
11	95.3	0.4	-39.13	-39.8
12 (100% bioC)	98.9	0.5	-39.20	-40.7

0.7% in this study. This is the minimal (1 σ) error in calculated bioC fractions as determined by the ¹⁴C method.

Cases 1 and 2 show the results for biogases of group 1 and cases 3 and 4 represent wood-based and landfill biogases (group 4). In these cases, the uncertainty in the approximated ${}^{14}a_{N(100\%bioC)}^{S}$ values is ± 0.5 pMC for cases 1 and 2 and ± 4 pMC for cases 3 and 4 (using Table 4). The possible deviation in

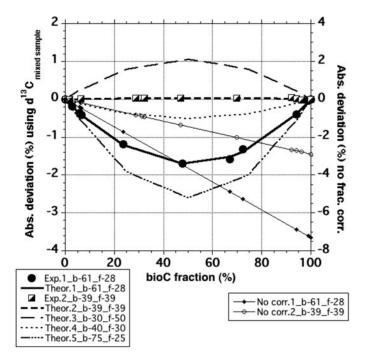


Figure 2 Absolute deviations in the calculated bioC fraction (%) for two experimental (exp.) and five theoretical (theor.) cases in which the measured $^{14}\mathrm{C}$ value has been corrected for isotope fractionation based on the $\delta^{13}\mathrm{C}$ value of the biofossil sample instead of the $\delta^{13}\mathrm{C}$ value of the bioC fraction in the sample alone. The figure also shows the absolute deviation in the bioC fraction for two cases in which no isotope fractionation correction has been applied at all (No corr.). In the figure text, b-61_f-28 means a $^{13}\mathrm{C}$ value of -61% for the bioC fraction and $^{13}\mathrm{C}$ value of -28% for the fossilC fraction.

the calculated bioC fraction due to incorrect isotope fractionation (if the δ^{13} C values of the separate bioC and fossilC fractions in the sample are unknown) varies between 0 (δ^{13} C_{bioC} = δ^{13} C_{fossilC}; cases 1 and 3) and a certain expected maximum value that depends on the possible difference between δ^{13} C_{bioC} and δ^{13} C_{fossilC} for the investigated type of sample material (δ^{13} C_{bioC} $\neq \delta^{13}$ C_{fossilC}; cases 2 and 4). The differences in δ^{13} C values between the biogenic and fossil gases investigated in this study range from +10‰ to -33‰. Differences up to 50‰ may be possible, but these differences will not occur often. Differences up to +40‰ and -40‰ are more likely and both these values have therefore been used in the calculation of the uncertainty range for cases 2 and 4. Cases 1 and 3 represent the minimal uncertainty in the calculated bioC fraction for biofossil gas mixtures for groups 1 and 4, respectively. For biogases with harvest years <2 yr or <10 yr before digestion (in Table 3: groups 2 and 3, respectively), the results are in between those of groups 1 and 4.

For verification whether biogases (especially the methane fraction) contain 100% bioC, the 100% bioC fraction can be determined with an (1σ) uncertainty of $\pm 0.7\%$, $\pm 0.9\%$, ± 1.7 , and $\pm 4\%$ for the biogas groups 1, 2, 3, and 4, respectively (Table 3). For biofossil gas mixtures, the (partly 1σ) errors in the calculated bioC fractions can be up to $\pm 2.5\%$ (at bioC of 50%), $\pm 2.5\%$ (at 55% bioC), $\pm 3.0\%$ (at 60% bioC), and $\pm 4.5\%$ (at 75% bioC) for the biogas groups 1, 2, 3, and 4 respectively.

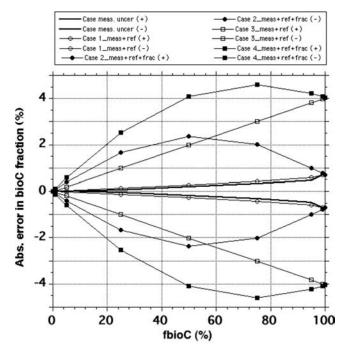


Figure 3 Ranges of possible (±) errors in the calculated bioC fraction (%), which are typical for specific biogas and biofossil gas mixtures if measured with AMS. Cases 1 and 2 represent biofossil gas mixtures with biogas from group 1 (as defined in Table 3). Cases 3 and 4 represent mixtures with biogas from group 4. In the figure text: "meas" is measurement uncertainty, "ref" is the uncertainty in the estimated 100% bioC reference value, and "frac" is the the systematic error in the isotope fractionation correction.

CONCLUSIONS AND RECOMMENDATIONS

This study measured the 13 C (δ^{13} C) values and 14 C ($^{14}a_N^S$) values of eight different biogas samples, three different natural gas samples and several biofossil gas mixtures with IRMS and AMS. The measured δ^{13} C values varied between -6% and +31% for the CO₂ fractions of the biogases, between -28% and -62% for the combusted CH₄ fractions of the biogases, and between -28% and -40% for the combusted natural gases. 14 C values of the investigated biogases varied between 102 and 116 pMC and were <0.2 pMC for natural gases.

A method is proposed to minimize the uncertainty in the calculated bioC fraction of a biogas or biofossil gas mixture, by dividing biogases into 4 different groups with different uncertainty ranges. This method can be used for other biofuels as well and we recommend its adoption in international standards. The uncertainty in the calculated bioC fraction due to the uncertainty in the approximated 100% bioC reference value ($^{14}a_{N(100\%bioC)}^{S}$) varies between $\pm 0.5\%$ and $\pm 3.5\%$ for the defined biogas groups.

Systematic deviations arise in the calculated bioC fractions if in the isotope fractionation correction the δ^{13} C value of the mixed biofossil sample is used instead of the δ^{13} C value of the bioC fraction alone. The size of this deviation increases with increasing difference between the δ^{13} C values of the bioC and fossilC fractions in the sample. For biofossil gas mixtures, differences can be up to 50% and deviations in the calculated bioC fraction of up to $\pm 2.5\%$ (at 50% bioC fraction) are therefore possible. If no isotope fractionation correction is applied in the calculation of the bioC fraction for biogases and biofossil gas mixtures, the absolute error in the determined bioC fraction can be up to -8%.

The possible errors in the determined bioC fractions were determined for the four biogas groups (Table 3). These values represent samples that have been measured with AMS, for which an approximated 100% bioC reference value has been used, and the δ^{13} C values of the separate biofossil fractions are unknown. For 100% bioC verification of biogases (biomethane), the 100% bioC fraction can be determined with an (1 σ) uncertainty of $\pm 0.7\%$, $\pm 0.9\%$, $\pm 1.7\%$, and $\pm 4\%$ for the biogas groups 1, 2, 3, and 4, respectively.

For biofossil gas mixtures, the errors in the calculated bioC fractions can be up to $\pm 2.5\%$ (at bioC of 50%), $\pm 2.5\%$ (at 55% bioC), $\pm 3.0\%$ (at 60% bioC), and $\pm 4.5\%$ (at 75% bioC) for the biogas groups 1, 2, 3, and 4, respectively. Knowledge about the origin and composition of the organic material used in the production of biogas is essential to minimize the uncertainties in the 14 C-based determination of the bioC fraction of biogas samples and biofossil gas mixtures.

ACKNOWLEDGMENTS

This project has been part of the research program EDGaR, which is partly funded by the contribution of The Northern Netherlands Provinces (SNN), European Union, European Fund for Regional Development, Ministry of Economic Affairs, and the Province of Groningen. DNV KEMA, in particular Klaas Bos and Harm Vlap are acknowledged for their help with obtaining different biogas and natural gas samples. The technical staff of the CIO involved in measuring the ¹³C and ¹⁴C values of the different samples, Henk Jansen, Bert Kers, Trea van Dijk-Dijkstra, Fsaha Ghebru, Dicky van Zonneveld, Henk Been, and Marc Bleeker, are thanked for their contributions.

REFERENCES

- European Commission. 2003. 2003/87/EC. Establishing a scheme for greenhouse gas emission allowance trading within the Community and Amending Council Directive 96/61/EC, Directive of the European Parliament and of the Council, Official Journal of the European Union, L275, 25.10.2003. p 32–46.
- Aerts-Bijma AT, Meijer HAJ, van der Plicht J. 1997.
 AMS sample handling in Groningen. Nuclear Instruments and Methods in Physics Research B 123(1-4):221-5.
- ASTM. 2012. ASTM D6866-12. Standard test methods for determining the biobased content of solid, liq-

- uid and gaseous using radiocarbon analysis. ASTM International, West Conshohocken, Pennsylvania, USA
- Bergamaschi P, Lubina C, Königstedt R, Fisher H. 1998. Stable isotopic signatures (δ¹³C, δD) of methane from European landfill sites. *Journal of Geophysical Research* 103(D7):8251–65.
- Cabral AR, Capanema MA, Gebert J, Moreira JF, Jugnia LB. 2010. Quantifying microbial methane oxidation efficiencies in two experimental landfill biocovers using stable isotopes. Water, Air, & Soil Pollution 209(1–4):157–72.

- Conrad R. 2005. Quantification of methanogenic pathways using stable isotopic signatures: a review and a proposal. *Organic Geochemistry* 36(5):739–52.
- de Rooij M, van der Plicht J, Meijer HAJ. 2008. Sample dilution for AMS 14 C analysis of small samples (30–150 μ g C). *Radiocarbon* 50(3):413–36.
- Dijs IJ, van der Windt E, Kaihola L, van der Borg K. 2006. Quantitative determination by ¹⁴C analysis of the biological component in fuels. *Radiocarbon* 48(3):315–23.
- Fellner J, Rechberger H. 2009. Abundance of ¹⁴C in biomass fractions of wastes and solid recovered fuels. *Waste Management* 29(5):1495–503.
- Fuex AN. 1977. The use of stable carbon isotopes in hydrocarbon exploration. *Journal of Geochemical Exploration* 7:155–88.
- Gonfiantini R. 1984. Stable Isotope Reference Samples for Geochemical and Hydrological Investigations. Vienna: IAEA. 77 p.
- Hsueh DY, Krakauer NY, Randerson JT, Xu X, Trumbore SE, Southon JR. 2007. Regional patterns of radiocarbon and fossil fuel-derived CO₂ in surface air across North America. *Geophysical Research Letters* 34: L02816, doi:10.1029/2006GL027032.
- Hua Q, Barbetti M, Rakowski AZ. 2013. Atmospheric radiocarbon for the period 1950–2010. *Radiocarbon* 55(4):2059–72.
- Hut G, Begemann MJS, Weerkamp HR. 1984. Determination of isotope ratios in the natural gas components CH₄ and N₂ separated by gas chromatography. Chemical Geology 46(1):75–83.
- ISO. 2012. ISO/FDIS 13833. Stationary source emissions Determination of the ratio of biomass (biogenic) and fossil-derived carbon dioxide Radiocarbon sampling and determination. [This standard is still under construction.] ISO, Geneva, Switzerland.
- Krzycki JA, Kenealy WR, DeNiro MJ, Zeikus JG. 1987. Stable carbon isotope fractionation by *Methanosar-cina barkeri* during methanogenesis from acetate, methanol, or carbon dioxide-hydrogen. *Applied and Environmental Microbiology* 53(10):2597–9.
- Laukenmann S, Polag D, Heuwinkel H, Greule M, Gronauer A, Lelieveld J, Keppler E. 2010. Identification of methanogenic pathways in anaerobic digesters using stable carbon isotopes. *Engineering in Life Sciences* 10(6):509–14.
- Levin I, Kromer B. 2004. The tropospheric ¹⁴CO₂ level in mid-latitudes of the Northern Hemisphere (1959–2003). *Radiocarbon* 46(3):1261–72.
- Levin I, Bergamanschi P, Dörr H, Trapp D. 1993. Stable isotopic signature of methane from major sources in Germany. Chemosphere 26(1–4):161–77.
- Levin I, Graul R, Trivett NBA. 1995. Long-term observations of atmospheric CO₂ and carbon isotopes at continental sites in Germany. *Tellus B* 47(1–2):23–34.
- Levin I, Kromer B, Schmidt M, Sartorius H. 2003. A novel approach for independent budgeting of fossil fuels CO₂ over Europe by ¹⁴CO₂ observations. *Geophysical Research Letters* 30(23):2194.

- Mann WB. 1983. An international reference material for radiocarbon dating. *Radiocarbon* 25(2):519–27.
- Meijer HAJ, van der Plicht J, Gislefoss JS, Nydal R. 1995. Comparing long-term atmospheric ¹⁴C and ³H records near Groningen, The Netherlands with Fruholmen, Norway and Izaña, Canary Islands ¹⁴C stations. *Radiocarbon* 37(1):39–50.
- Milkov AV. 2005. Molecular and stable isotope compositions of natural gas hydrates: a revised global dataset and basic interpretations in the context of geological settings. *Organic Geochemistry* 36(5):681–702.
- Mohn J, Szidat S, Fellner J, Rechberger H, Quartier R, Buchmann B, Emmenegger L. 2008. Determination of biogenic and fossil CO₂ emitted by waste incineration based on ¹⁴CO₂ and mass balances. *Bioresource Technology* 99(14):6471–9.
- Mohn J, Szidat S, Zeyer K, Emmenegger L. 2012. Fossil and biogenic CO₂ from waste incineration based on a yearlong radiocarbon study. Waste Management 32(8):1516–20.
- Mook WG. 1980. The effect of fossil fuel and biogenic CO₂ on the ¹³C and ¹⁴C content of atmospheric carbon dioxide. *Radiocarbon* 22(2):392–7.
- Mook WG. 2000. Environmental Isotopes in the Hydrological Cycle. Volume 1, Introduction, Principles, Methods. Vienna: UNESCO/IAEA. 255 p.
- Mook WG, van der Plicht J. 1999. Reporting ¹⁴C activities and concentrations. *Radiocarbon* 41(3):227–39.
- Norton GA, Devlin SL. 2006. Determining the modern carbon content of biobased products using radiocarbon analysis. *Bioresource Technology* 97(16):2084– 90.
- Palstra SWL, Meijer HAJ. 2010. Carbon-14 based determination of the biogenic fraction of industrial CO₂ emissions application and validation. *Bioresource Technology* 101(10):3702–10.
- Palstra SWL, Karstens U, Streurman H-J, Meijer HAJ. 2008. Wine ethanol ¹⁴C as a tracer for fossil fuel CO₂ emissions in Europe: measurements and model comparison. *Journal of Geophysical Research* 113: D211302, doi:10.1029/2008JD010282.
- Rosenfeld WD, Silverman SR. 1959. Carbon isotope fractionation in bacterial production of methane. *Science* 130(3389):1658–9.
- Schoell M. 1980. The hydrogen and carbon isotopic composition of methane from natural gases of various origins. *Geochimica et Cosmochimica Acta* 44(5):649–61.
- Staber W, Flamme S, Fellner J. 2008. Methods for determining the biomass content of waste. Waste Management & Research 26(1):78–87.
- Stuiver M. 1983. International agreements and the use of the new oxalic acid standard. *Radiocarbon* 25(2):793–5.
- Stuiver M, Polach HA. 1977. Discussion: reporting of ¹⁴C data. *Radiocarbon* 19(3):355–63.
- van der Plicht J, Wijma S, Aerts AT, Pertuisot MH, Meijer HAJ. 2000. Status report: the Groningen AMS facility. Nuclear Instruments and Methods in Physics Research B 172(1–4):58–65.

- Weiland P. 2010. Biogas production: current state and perspectives. *Applied Microbiology and Biotechnology* 85(4):849–60.
- Whiticar MJ, Faber E, Schoell M. 1986. Biogenic methane formation in marine and freshwater environments: CO₂ reduction vs. acetate fermentation—isotope evidence. *Geochimica et Cosmochimica Acta* 50(5):693–709.
- Wijma S, Aerts AT, van der Plicht J, Zondervan A. 1996. The Groningen AMS facility. *Nuclear Instruments*
- and Methods in Physics Research B 113(1-4):465-9
- Zyakun AM, Muravyev AI, Baskunov BP, Laurinavichius KS, Zakharchenko VN, Peshenko VP, Lykov IN, Shestakova GA. 2010. Estimation of microbial methane generation and oxidation rates in the municipal solid waste landfill of Kaluga city, Russia. *Isotopes in Environmental and Health Studies* 46(1):78–90.