

Forage kochia seed germination response to storage time and temperature

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Abstract

The Eurasian low-shrub, forage kochia [*Kochia prostrata* (L.) Shad.], was introduced into western North America for use in restoration of severely disturbed landscapes in arid and semi-arid environments. Seed mature in late fall and are short-lived in typical warehouse conditions. In a preliminary, cold-temperature experiment (2° C) using 3-month-old seed from 16 forage kochia accessions, mean germination time, expressed as days to 50% germination, varied from 4 to 88 days. Follow up experiments using seed of 5 accessions tested the effects of storage time and temperature on seed viability and mean germination time and related this to field planting success. Sub-samples were air-dried and stored in plastic bags in a freezer, cold room, and lab (–15, 2, and 20° C respectively). A fourth set of subsamples was stored in a shed with no temperature control (simulated warehouse storage). Seed were tested fresh and retested after 4, 8, 12, 24, and 36 months of storage. Mean viability decreased from 77% (range 66 to 93%) for recently harvested seed, to 24 and 8% for lab- and shed-stored seed, after 36 months of storage. No significant change in viability was observed for cold room- and freezer-stored seed. Across all accessions, cold temperature mean germination time (MGT) for recently harvested seed was 73 days (range 51 to 109 days). For each accession, germination occurred primarily over a 70 day period. Mean germination time decreased as storage time increased for lab- and shed-stored seed, varied unpredictably for cold room-stored seed, and remained unchanged for freezer-stored seed. Field germination using 1- and 2-year old lab- and shed-stored seed was significantly faster than that of same-aged cold room- and freezer-stored seed. The number of live seedlings 4 months after planting for cold room- and freezer-stored seed was 10-fold or greater than that of lab- and shed-stored seed. Thus a delayed, asynchronous cold-temperature germination pattern appears to be adaptive for forage kochia establishment. Cold, dry storage prevents loss of seed viability and preserves this desirable germination pattern.

Key Words: *Kochia prostrata*, prostrate summer cypress, germination rate, mean germination time, after-ripening, germination synchronization, cold-desert revegetation

Resumen

El arbusto Euroasiático “Kochia forrajera” [*Kochia prostrata* (L.) Shad], se introdujo en oeste de Norteamérica para usarlo en la restauración de paisajes severamente degradados de ambientes áridos y semiáridos. La semilla madura a finales de otoño y bajo las condiciones típicas de almacenamiento en bodega tiene una vida corta. En un experimento preeliminar, utilizando bajas temperaturas (2° C), semilla de 3 meses de cosechada y proveniente de 16 entradas de “Kochia forrajera”, se estableció que el tiempo promedio de germinación, expresado como días al 50% de semillas germinadas, varió de 4 a 88 días. En experimentos subsecuentes, usando semilla de 5 entradas, probamos los efectos del tiempo de almacenaje y la temperatura en la viabilidad de la semilla y el promedio del tiempo de germinación, y relacionamos esto con el éxito de establecimiento de las plantas en el campo. Submuestras se secaron con aire, se guardaron en bolsas de plástico y se almacenaron en: 1) congelador, 2) cuarto frío y 3) laboratorio (–15, 2 y 20° C respectivamente). Un cuarto grupo de submuestras se almacenó en un cobertizo sin control de temperatura (simulando las condiciones de bodega). Las semillas se evaluaron recién cosechadas y se reevaluaron a los 4, 8, 12, 24, y 36 meses de almacenamiento. El promedio de viabilidad disminuyó de 77% (rango de 66 a 93%) en semillas recién cosechadas a 24 y 8% en semillas almacenadas durante 36 meses en laboratorio y cobertizo. No se observaron cambios significativos en la viabilidad de semillas almacenadas en congelador y cuarto frío. La media general de tiempo de germinación en temperaturas frías de todas de las entradas de “Kochia forrajera” fue de 73 días en semillas recién cosechadas (rango de 51 a 109 días). Para cada entrada de “Kochia forrajera” la germinación ocurrió después de 70 días. El tiempo promedio de germinación de semillas almacenadas en el laboratorio y cobertizo disminuyó conforme el tiempo de almacenamiento aumentó, varió en forma impredecible para la semilla almacenada en cuarto frío y permaneció sin cambios para la semilla almacenada en congelador. La germinación en campo de semillas almacenadas por 1 y 2 años en el laboratorio y cobertizo fue significativamente más rápida que la de semilla almacenada durante el mismo tiempo en cuarto frío o congelador. El número de plántulas vivas (4 meses después de la siembra) de semillas almacenadas en cuarto frío y congelador fue 10 veces o más que el de semillas almacenadas en laboratorio y cobertizo. Por lo tanto, un patrón de germinación retasado asincrónico de temperatura fría parece ser que se adapta para el establecimiento de “Kochia forrajera”. El almacenamiento en condiciones secas y frías previene la pérdida de viabilidad y conserva este patrón deseable de germinación.

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Many arid and semiarid rangelands of the western United States are becoming destabilized due to the interrelated effects of unsustainable grazing practices (Young 1994), introductions of old-world weeds (Peters and Bunting 1994), and changes in wildfire frequency (Whisenant 1990). Unchecked, these forces are causing the conversion of shrub and perennial grasses-dominated communities to annuals-dominated communities with reduced resource values and serious management considerations. Restoring function and desired use to degraded rangelands can be accomplished by seeding adapted perennials capable of competing with weedy species and resprouting after fire. The need to develop plant materials endowed with these traits and the capacity to coexist with and complement remnant populations of native communities, is widely recognized.

Forage kochia [*Kochia prostrata* (L.) Shrad.], also known as prostrate summer cypress, is a polymorphic low-shrub native from the Mediterranean Basin to Siberia (Shishkin 1936). Baylan (1972) recognizes 2 subspecies: sp. *virescens* (Frenzl) Prat., commonly known as green-stem forage kochia, and sp. *grisea* Prat., or gray-stem forage kochia. Several germplasm introductions have been made to evaluate the utility of forage kochia for revegetation of severely disturbed arid and semi-arid sites in the western United States (McArthur et al. 1974, Frischknecht and Ferguson 1984, Blauer et al. 1993). Introductions have been evaluated for such traits as livestock and wildlife preference (Davis and Welch 1985), nutritional quality (Davis and Welch 1983, 1985), seed germination characteristics (Young et al. 1981, Waller et al. 1983, Briede and McKell 1992, Stewart 1998), establishment and competitive attributes (Stevens and Van Epps 1984, McArthur et al. 1990, Monsen and Turnipseed 1990, Stevens and McArthur 1990, McArthur et al. 1996, Harrison et al. 2000), and salinity tolerance (Francois 1976, Romo and Haferkamp 1987). Studies demonstrate that forage kochia is well suited for a variety of soil types on cold-desert rangelands receiving 150–300 mm annual precipitation. A single cultivar, 'Immigrant', (sp. *virescens*) was released after demonstrating wide spread adaptability, forage quality, and productivity (Stevens et al. 1985). Several thousand kilograms of seed of this cultivar are planted on rangelands most years.

Forage kochia grows throughout the summer, flowering indeterminately from June through September (Shishkin 1936, Baylan 1972). Fruits usually mature by the

middle of October (Waller et al. 1983, Stewart 1998) and are commonly harvested from October through January. Subsequently, current-year seed is sometimes not available in time for fall and early winter revegetation projects. Forage kochia stand establishment from seed stored for 1 or more years in typical warehouse conditions is generally poor. This is due in part to the short shelf-life for seed of this species. Shelf-life can be extended by adequate pre-drying and by storage in sealed containers at low temperatures (Young et al. 1981, Jorgensen and Davis 1984, Stewart 1998). However, a lack of stand establishment success using high-viability, stored seeds is more difficult to explain. Low seed vigor has been proposed as one possible explanation (Haferkamp et al. 1990). In this work, we provide evidence to support an alternative explanation.

For best emergence, forage kochia seed must germinate at or near the soil surface. In the cold-desert environments to which this species is adapted, soil temperature and moisture conditions favorable for surface germination and seedling growth occur most dependably during brief periods in late winter and early spring. Seed that germinate at other times (e.g. too late or too early) have a higher risk of mortality than do seed with proper germination timing. Therefore, we attempt to test the

hypothesis that seeding failure using stored but viable seed may be caused by storage-related alteration of germination timing controls.

Our objectives were to examine in the laboratory among-accession differences in cold-temperature germination rate, or cold mean germination time, expressed as days to 50% germination (based on total viable seeds), and determine the effects of storage time and temperature on seed viability and mean germination time for forage kochia. We also relate results of field plantings using differently-stored seed to test conclusions drawn from laboratory experiments.

Materials and Methods

A preliminary experiment was conducted to determine among-accession differences in mean germination time for forage kochia. Three-month-old seed from 15 experimental lines collected from a common garden located near Boise, Ida. (Latitude and Longitude; 43° 20' North 116° 35' West) were studied. All accessions except 1 (origin Peoples Republic of China, Table 1) were sp. *grisea*. Seed were hand-collected, dried, and cleaned in November 1989. In addition, because it

Table 1. Forage kochia accessions by plant introduction number, location and soil type of original collection, and cold temperature (2° C) mean germination time (MGT), expressed as days to 50% germination of viable seeds. All seed was 3 months old except for accession 314929 which was 1 year old. Means followed by the same letter are not significantly different at the $p < 0.05$ level (Tukey).

P.I. No. ^a	Original Collection Information ^b		
	Location	Soil Type	MGT (days)
-----	Inner Mongolia, Peoples Republic of China	Unknown	3.7j
314929 ^{c,d}	Stavropol', Russia "Immigrant"	Sandy	11.2j
330708	Rashat, Iran	Unknown	41.9hi
343101 ^d	Kyrgyzstan	Unknown	32.1hi
356817	Akt'ubinsk, Kazakhstan	Salty	44.7gh
356818 ^d	Akt'ubinsk, Kazakhstan	Clay	41.5hi
356819	Akt'ubinsk, Kazakhstan	Salty	48.7efgh
356820	Akt'ubinsk, Kazakhstan	Sandy	53.1defg
356821	Akt'ubinsk, Kazakhstan	Salty	50.9efgh
356822	Akt'ubinsk, Kazakhstan	Clay	55.6cdef
356823	Akt'ubinsk, Kazakhstan	Sandy	58.5bcde
356824	Akt'ubinsk, Kazakhstan	Salty	67.0b
356825	Akt'ubinsk, Kazakhstan	Sandy	66.4bc
356826 ^d	Akt'ubinsk, Kazakhstan	Salty	88.3a
358941 ^d	Stavropol', Russia	Sandy	47.6fgh
422519	Unknown	Unknown	61.2bcd

^aThe PI number is the USDA plant inventory number. The first 2 accessions are sp. *virescens* and the rest belong to sp. *grisea*.

^bThis information came with original seed packets from the USDA ARS Western Regional Plant Introduction Station in Pullman, Washington. Locations should be taken as general areas. The collections from Akt'ubinsk are regional collections taken from the Aral Sea to the Ural Mountains. Those from Stavropol' came from a botanical garden.

^cThe cultivar 'Immigrant' (Stevens et al. 1985).

^dAccessions selected for the storage experiments.

had not been collected at the Boise common garden, 1-year-old seed of Immigrant forage kochia (sp. *virescens*) was secured and included in the experiment. For each accession, 4 replications of 50 seeds each were placed in 15 x 100 mm plastic petri dishes on top of 2 germination blotters (Anchor Paper Company, St. Paul, Minn.) moistened to saturation with tap water. Petri dishes were arranged randomly in a cardboard box. The box was enclosed in a plastic bag to retard desiccation, and placed in a walk-in cold room at 2° C. Water was added periodically to maintain blotter water content at near saturation levels. Seed were examined and germinants were counted and removed from the petri dishes on a weekly basis for 112 days. Seed were classified as germinated when radicles had elongated at least 3 mm and demonstrated a positive gravitropic response. Most germinants were also at least partially uncoiled. At the end of 112 days, the box was moved to a germination chamber set at 15/25° C (12 hour alternating) for 3 additional days. After final germination counts were made, an embryo integrity (squish) test confirmed the absence of remaining viable (dormant) seed. Viability percentages and mean germination time values were calculated for each replication.

Storage Experiments

Seed of 4 experimental lines were harvested from a nursery site in Spanish Fork, Utah (Latitude and Longitude; 39° 40' North 111° 5' West) during October and November 1992 (Table 1). Length of harvest period was due to variability in ripening rates among accessions. Ripened fruits were easily collected by beating fruit-bearing stems against the edge of hand-held hoppers with badminton rackets. Harvested fruits were spread on tables to a depth of 10 cm for 10+ days prior to further processing. Temperatures varied between 15 and 25° C, and though not measured, relative humidity was sufficiently low to allow drying to occur. Subsequently, inert material and a portion of unfilled fruits were removed from each lot using a 2-screen fanning mill. Cleaned seed (fruits) were stored in cloth bags for approximately 1 month (15 to 25° C) before further processing. A fifth accession, non-certified Immigrant, was acquired from a commercial source and had been collected in November 1992 from a USDI-BLM planting located 15 km northeast of Milford, Utah.

In December, seed of all 5 accessions were oven-dried at 35° C for 72 hours and

subdivided into 4 equal sublots. Each subplot was placed in a labeled plastic bag closed with a rubber band. One subplot (bag) for each accession was placed inside each of 4 larger plastic bags, also closed with a rubber band. One of each of these bags was stored for 36 months in: 1) the laboratory at room temperature (20° C); 2) the walk-in cold room (2° C); 3) a freezer (-15° C); and 4) a shed in Provo, Utah with no temperature control (to simulate warehouse storage). To determine seed water content immediately after processing and again after 8 months of storage, subsamples of approximately 20 g were weighed, oven dried for 24 hours at 65° C, and reweighed. Mean water content for dried seed before storage was 4.1%. There was no significant change in water content after 8 months of storage for all storage treatments (mean = 3.9%).

Germination procedures followed those used in the preliminary experiment with the following exceptions: 1) 25 instead of 50 seeds per replication were used and 2) final incubation temperature was 10/20° C instead of 15/25° C. These changes were intended to make the germination environment less favorable for potential seed pathogens. Each accession was tested fresh (December 1992) and each accession/storage combination was tested after 4, 8, 12, 24, and 36 months of storage.

Field Trials

A fallowed agricultural field with uniform soil and topography was selected near American Fork, Utah (Latitude and Longitude; 40° 20' North 111° 50' West; elev. 1,380 m) as a site for field germination and seedling establishment experiments. The site was tilled each fall prior to planting dates (30 November 1993 and 2 December 1994). Litter cover was less than 5%, ensuring good seed contact with mineral soil. Results were interpreted in light of ambient temperature data for the Utah Lake Lehi weather station (NOAA 1993, 1994, 1995) located 8 km west of the planting site (same approximate elevation). When data were incomplete for any month, estimates were made using data from the Vernon and Timpanogos Cave sites, 48 and 8 km distant from the planting site.

Only Immigrant seed was used for these field studies. Seed had experienced 1 and 2 years of storage prior to planting. In preparation, each year twelve, 0.2-g and three, 2-g subsamples were weighed from each of the 4 Immigrant sublots (storage treatments). The twelve, 0.2-g subsamples of approximately 160 seeds each (150

viable seeds prior to storage) were enclosed in marked 10 x 10 cm square nylon-mesh bags. Bags were randomly sorted into 12 sets, each set including 1 bag from each storage treatment.

In 1993, each set of bags was buried under 1.0 to 1.5 cm of soil and marked with a 75-cm survey flag to facilitate detection in snow. Each group of bags was placed under a 20 cm (diameter) wire-mesh cone buried 3 cm deep to discourage rodents. Cones were arranged in a 2 by 6 grid with 10–20 cm inter-cone spacing. We used similar procedures in 1994, however, bags were not buried in soil and cones were not used. Instead, bags were firmly pressed into the soil surface and covered with 1–2 cm of straw. This change was made to simplify extraction of bags from frozen soil. This mulch was held in place by 2 criss-crossed layers of pine stakes (2 x 4 x 35 cm) placed horizontally with 12 to 15 cm spacing between stakes. No rodent predation was encountered either year.

Each year, 3 sets (replications) of 4 bags each were randomly selected and retrieved from the site 1 and 2 months following burial. Retrieved bags were packed in loose snow and taken directly to the laboratory for processing. There, soil and ice were gently washed from each bag before opening. We then opened bags and determined germination status for each seed. Average time from bag retrieval to final seed evaluation was 2 to 3 hours. Due to rapid germination associated with unseasonably warm temperatures in December and January (Fig. 1), 6 sets of bags remained unused both years.

The 2-g samples were planted on twelve, 1.0 x 0.5 m plots arranged in a randomized block design each year. Planting consisted of hand broadcasting seed onto the appropriate plot and lightly raking over the entire plot to improve soil/seed surface contact and to anchor the seeds to the plots. Planting dates were the same as burial dates for the retrieval experiments. We determined the number of forage kochia seedlings per plot approximately 4 months after planting (early April), both years. At this time, the soil surface was dry and crusted and surviving seedlings had reached or passed the 6-leaf stage.

Percentage data (seed viability in storage experiments and seed germination and seedling survival in field experiments) were arcsine transformed for statistical analysis. Results were analyzed using the GLM procedure (SAS 1998) to test for effects of storage treatment and time on

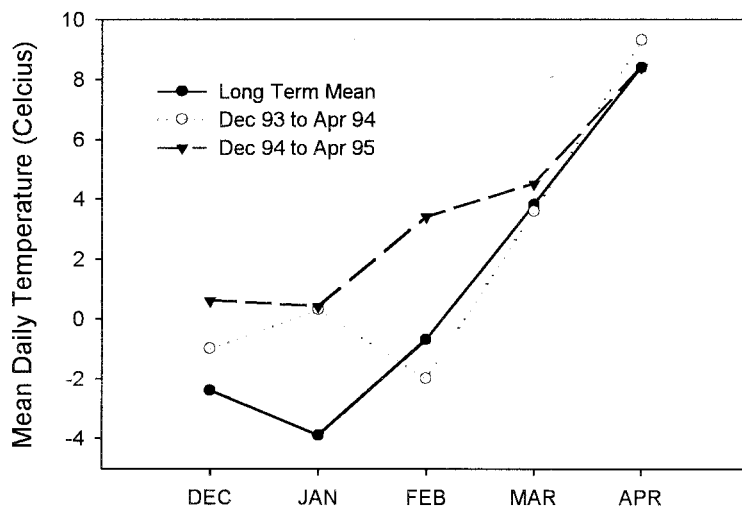


Fig. 1. Mean daily temperatures for the months of December to April for the Utah Lake Lehi weather station. Values for the winters of 1993–1994 and 1994–1995 are compared to the long-term mean (1961–1990) for the site (NOAA 1993, 1994, 1995). When data were missing, means were estimated using data from the Vernon and Timpanogos Cave stations.

viability percentage and cold mean germination time in laboratory experiments, and field germination and seedling survival in field experiments. Significant differences among accession and treatment means were determined using the Tukey-Kramer (Tukey) multiple means comparison test.

Results

Mean germination time (MGT) for the 15 experimental lines of the preliminary experiment ranged from 3.7 to 88.3 days (Table 1). The fastest germination rates belonged to the accession originally collected in the Inner-Mongolia region of the

People's Republic of China (MGT = 3.7 days) and to 1 year-old Immigrant seed (MGT = 11.2 days). Mean germination time for the remaining accessions (all recently harvested seed) was 54.1 days. Maximum percentage of ungerminated seed after 112 days of chilling was 29% (MGT = 88.3 days). Accessions selected for the storage experiments represented nearly the full range in observed mean germination time values.

Storage Experiments

Mean viability of recently harvested seed was 77% (range 66 to 93%). After 12 months storage, viability of lab- and shed-stored seed (71%) was significantly lower

($p < 0.05$) than that of cold room- and freezer-stored seed (83%). After 36 months of storage, viability of lab- and shed-stored seed decreased to 24 and 8%, respectively (Fig. 2). Viability percentages for 36-month old cold room- and freezer-stored seed were not significantly different than those of recently harvested seed. However, viability of 24-month old cold room- and freezer-stored seed was significantly lower than that of either recently harvested or 36-month old seed for 4 of 5 accessions. Because viability can not increase through time, the observed increase is best explained as differential selection among test dates for filled fruits. This is not surprising in as much as fill for intact fruits can be difficult to detect. Only with the commercially collected and conditioned Immigrant seed were we able to consistently eliminate most empty fruits by visual inspection. Within test dates, treatment-related differences in viability estimates were similar for all accessions (including Immigrant), suggesting that within-test date seed selection was much more consistent than among-test date selection. All ungerminated, viable seed germinated within a few days after exposure to warmer germination temperatures (10/20° C) at the end of the 112-day cold treatment.

Although similar in duration (approximately 70 days), the primary period of germination for recently harvested seed varied considerably among accessions (Fig. 3). Mean germination time (MGT) was 73 days (range = 51 to 109 days). As in the preliminary experiment, significant among-accession differences in germination rate were observed (Table 2). Mean germination times for these accessions were greater for this experiment than in the preliminary experiment, suggesting differences in afterripening and/or site conditions during seed maturation. This difference was most pronounced for seed of Immigrant (MGT = 11 vs. 70 days). However, Immigrant seed in the preliminary experiment was 1-year-old, therefore, comparisons between that seed and recently harvested seed of the storage experiment would be invalid. A more appropriate inter-experimental comparison for Immigrant seed germination rate would be between results of the preliminary experiment and those derived after 12 months of lab- or shed-storage (MGT = 13 and 15 days, respectively).

After 8 months of storage, mean germination times for lab-, shed-, and cold room-stored seed (17, 19, and 22 days, respectively) was significantly shorter

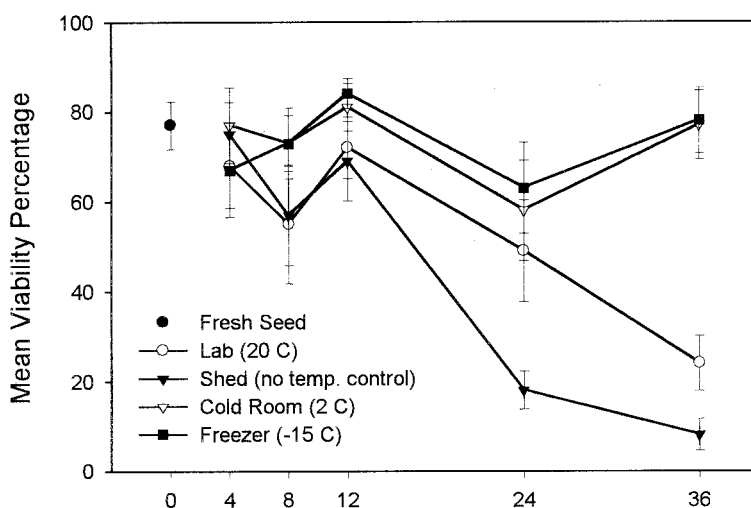


Fig. 2. Mean viability for 5 accessions of forage kochia tested fresh and in response to dry storage treatments.

Table 2. Cold temperature (2° C) mean germination time (days to 50% germination of viable seeds) for 5 accessions of forage kochia tested fresh and after 4-36 months dry storage in a lab (20° C), cold room (2° C), freezer (-15° C), and shed with no temperature control. Among accessions for fresh seed, and within accessions and storage period for stored seed, means followed by the same letter are not significantly different at the $p < 0.05$ level (Tukey).

Accession No./ Storage Treatment	Fresh	Mean Germination Time (days to 50% germination)				
		Storage Period (months)				
		4	8	12	24	36
----- (days) -----						
314929 (Immigrant)	70b					
Lab		47b	16b	13c	11b	6b
Cold Room		44b	26b	36b	58a	16b
Freezer		71a	70a	70a	67a	76a
Shed		59a	19b	15bc	21b	15b
343101	51c					
Lab		40a	12b	17b	12b	8b
Cold Room		21b	14b	24b	46a	28b
Freezer		54a	56a	64a	55a	59a
Shed		49a	17b	13b	12b	9b
356818	53c					
Lab		43ab	21b	23b	13b	16b
Cold Room		26b	19b	37b	51a	31b
Freezer		49a	49a	63a	58a	67a
Shed		59a	20b	16b	18b	19b
356826	109a					
Lab		61b	17b	7c	11b	6c
Cold Room		62b	31b	66a	86a	37b
Freezer		96a	93a	87a	90a	106a
Shed		78ab	21b	13b	10b	---
358941	81b					
Lab		55ab	20b	12b	12b	6b
Cold Room		40b	20b	23b	63a	8b
Freezer		69a	87a	67a	76a	75a
Shed		62a	18b	14b	12b	11b

than those observed for recently harvested seed (Fig. 4). Within-accession changes and among-accession differences in mean germination time were small and essentially insignificant for lab- and shed-stored seed in response to longer periods of storage. Mean germination time for cold room-stored seed (all 5 accessions) changed unpredictably over the duration of the experiment with values significantly higher ($p > 0.05$) after 12 and 24 months (37 and 61 days, respectively), but not after 36 months (24 days) when compared to those observed after 8 months (22 days) of storage. Across all storage periods, differences in the observed mean germination time for freezer-stored seed were not significantly different than that of fresh seed. Although initial (recently harvested seed) mean germination times varied more than 2-fold among accessions, relative differences in treatment effects are similar among accessions (Table 2). For all accessions, freezer storage was the only method found effective in preserving original germination patterns through 36 months of storage.

Field Trials

Laboratory germination results generally supported results observed for field experiments. For both years of burial, significantly higher percentages of viable seed had not yet germinated 1 month after planting for cold room- and freezer-stored seed than for lab- and shed-stored seed (Table 3). Nearly all viable seed from all 4 storage treatments had germinated by the second retrieval date. Unseasonably warm temperatures during December and January both years (1993–94 and 1994–95; Fig. 1) undoubtedly allowed germination to occur at a more rapid rate than might have been observed under

colder conditions.

Mean numbers of live seedlings counted on the 0.5 m² emergence plots for cold room- and freezer-stored seed were approximately 10-fold those counted for lab- and shed-stored seed the first year (Table 4). Differences in second-year results are greater than those of the first year. This is true whether expressed as the absolute number of seedlings counted or as a percentage of viable seed planted as estimated by retrieval experiments. Differences between cold room- and freezer-stored seed and between lab- and shed-stored seed were not significant either year.

Discussion

Forage kochia is well adapted to the sometimes severe and generally variable weather conditions found in the cold-deserts of western North-America (McArthur et al. 1974, Pendleton et al. 1992, McArthur et al. 1996). Although successful seedling establishment may require some disturbance in perennial communities (McArthur et al. 1990, Stevens and McArthur 1990, Harrison et al. 2000) new seedlings are observed near established plants most years. Late-season seed maturation and a delayed/asynchronous germination pattern in cold temperatures result in a high probability that some fraction of seed will germinate during a time favorable for establishment success. Germination that is too early or too uniform places seedlings at risk to conditions unfavorable for survival. The timing of these late winter/early spring 'windows of opportunity' varies annually, favoring the selection of a bet-hedging germination strategy (Phillipi and Seger 1989). Asynchronous cold-temperature germination functions as a bet-hedging strategy for forage kochia. Changes in seed germinability in dry storage, also known as dry afterripening, are common and are generally expressed as loss of innate dormancy, loss of light and temperature sensi-

Table 3. Field germination of Immigrant forage kochia seed 1 and 2 months after planting as affected by storage treatment. Seed planted in 1993 and 1994 had been stored dry for 1 and 2 years before planting. Data are shown as mean number of viable seed ungerminated and percent ungerminated based on total viable seed (in parentheses). Within rows, means followed by the same letter are not significantly different at the $p < 0.05$ level (Tukey).

Year of Planting	Months after Planting	Storage Treatment			
		Lab	Shed	Cold Room	Freezer
		No. (%)	No. (%)	No. (%)	No. (%)
93	1	60 (51)b	61 (49)b	138 (92)a	135 (92)a
	2	18 (15)ab	6 (8)b	26 (17)a	32 (21)a
94	1	3 (4)b	3 (22)b	113 (76)a	124 (84)a
	2	0b	0b	6 (4)ab	20 (7)a

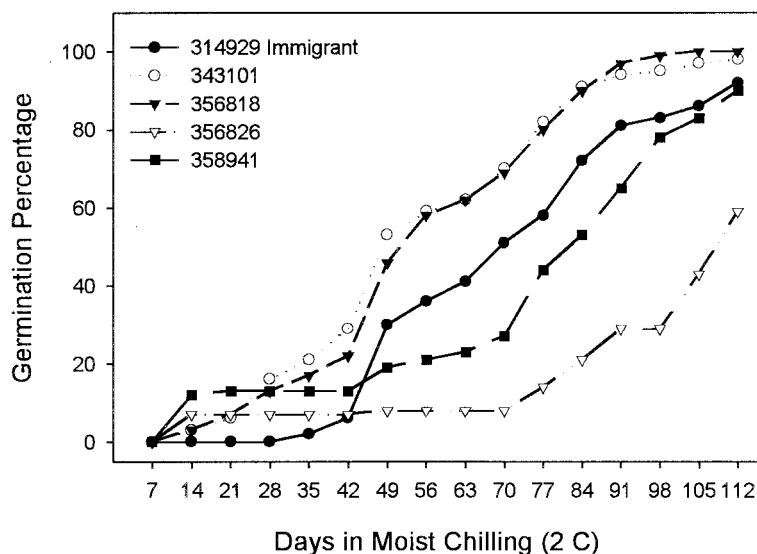


Fig. 3. Time course germination plots for 5 recently-harvested accessions of forage kochia at 2° C.

tivity, and/or changes in germination rate (Bewley and Black 1982). After-ripening is uni-directional and temperature dependent. Therefore, it can be expected that the process of after-ripening is greatly slowed, or even arrested, at freezing temperatures. Our data suggest that this is the case for forage kochia seed.

Conversely, we might expect seed stored at constant warm temperature to show a relatively rapid, continuous change in germinability, or in our case, decrease in mean germination time (MGT). Seed

stored in the lab for the first 8 months at room temperature followed just such a pattern. A leveling off of MGT after 8 months of storage suggests that warm-stored seed reached a threshold in germination rate. Temperatures for shed-stored seed were controlled by ambient temperatures which fluctuate daily and seasonally. For much of the first 4 months of storage (January–April) ambient temperatures were cold, often sub-freezing (Fig. 1), resulting in slower rates of afterripening. We might predict that 4-month MGT for

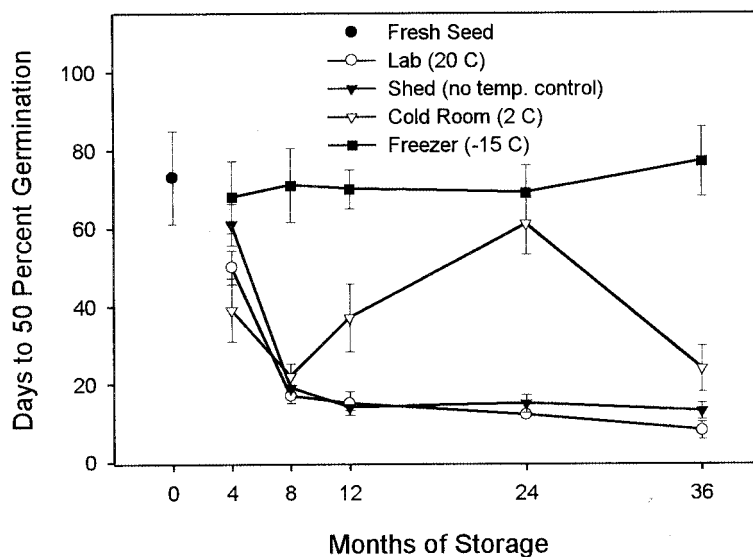


Fig. 4. Mean germination time (days to 50% germination) for 5 accessions of forage kochia as affected by storage location (temperature) and time.

this storage treatment would have to be intermediate to that of freezer- and lab-stored seed. This was indeed the case (Fig. 3). In the time period between 4 and 8 months of storage (May–August), seed in the shed storage treatment were exposed to ambient temperatures higher than in any other storage treatment. Subsequently, changes related to after-ripening should have been accelerated for this storage treatment during this time. Not surprisingly, the change in mean germination time for all accessions in the 4 to 8 month interval was greater for this storage treatment than for any other. From this point on, mean germination time for shed- and lab-stored seed did not differ significantly with length of storage.

Accounting for the changes in mean germination time (MGT) for cold room-stored seed over time is more difficult. Because after-ripening is temperature dependent, we expected changes in MGT for cold room-stored seed to be slower than those observed for lab-stored seed. This was not the case for the first 8 months of storage suggesting that another process, perhaps akin to prechill (stratification), may have been responsible. However, prechill treatments are generally effective only when seed have imbibed water. Our seed remained dry through the storage treatment. Waller et al. (1983) reported that after 3 months dry storage, forage kochia seed stored at 4° C produced germination percentages higher than seed stored at either -18 or 21° C, also suggesting an accelerated rate of after-ripening in near freezing temperatures. Increases in MGT associated with 12 and 24 months of storage and subsequent decrease observed with the 36 month treatment are puzzling and identifying a process or processes responsible for these unexpected reversals will not be easy.

Among accession differences in MGT for freshly collected seed were significant and likely adaptive, probably reflecting climatic differences in the Eurasian origins of the respective accessions. We have insufficient knowledge concerning those origins to speculate on the significance of those differences including the unusually rapid cold germination rate of 3-month old seed of the Chinese accession (Table 1). Waller et al. (1983) observed differences in germination (15/25° C) for 2 accessions of forage kochia (one of each subspecies) as affected by harvest date, drying method, and storage temperature. They concluded that, based on their data, the gray-stem variety (sp. *grisea*) had better germination characteristics than the red-

Table 4. Number of forage kochia seedlings 5 months after planting as affected by seed storage treatment. Seed planted in 1993 and 1994 had been stored for 1 and 2 years. Numbers in parentheses are live seedlings expressed as a percent of viable seed planted. Within rows, means followed by the same letter are not significantly different at the $p < 0.05$ level (Tukey).

Year of Planting	Storage Treatment			
	Lab	Shed	Cold Room	Freezer
	No. (%)	No. (%)	No. (%)	No. (%)
1993	30 (2)b	22 (2)b	240 (16)a	207 (14)a
1994	10 (1)b	2 (2)b	278 (20)a	292 (21)a

stem variety (sp. *virescens*) and suggest that it would therefore be preferred for revegetation. Our study significantly expands understanding of forage kochia intra-specific variability in germination behavior both because of the quantity of germplasm tested and because of the exposure of that germplasm to treatments relevant to storage and seedbed environments. We contend that the germination patterns for recently collected seeds of all accessions that we have tested (with the likely exception of the Chinese accession) are broad enough for successful establishment in the arid/semiarid West. Further speculation concerning how adaptive the germination timing of any one accession might be for specific environments, based on our data, would be inappropriate. Changes in MGT in storage were similar for all accessions even though initially values varied 2-fold. Ultimately, MGT for all accessions dropped to between 21 and 12 days after 8 months of lab or shed storage suggesting that similar storage treatments are unsuitable for any and all accessions to preserve a germination profile suitable for wildland plantings.

Storage treatment related differences in laboratory germination trials were largely matched by field seed retrieval experiments. In addition, successful seedling establishment was significantly greater for cold room- and freezer-stored seed; those treatments with the slowest MGT in laboratory experiments after 12 and 24 months (Table 2).

Conclusions

Data from this study support the conclusion that an altered germination timing regime may be responsible for forage kochia seeding failure using after-ripened seed of known high viability. We observed that low temperature germination is delayed and asynchronous for recently harvested seed and that germination rate increases dramatically with after-ripening. We also observed poor field performance of after-ripened seed. With this study we

provide additional evidence of why forage kochia seed need to be dried to less than 7% moisture, sealed in water tight containers, and stored at temperatures below 5° C (Jorgensen and Davis 1984, Stewart 1998) when not planted soon after harvest. Because of the unpredictable changes in germination patterns we observed for cold room-stored seeds, we believe that there may be an additional advantage to storage at temperatures below freezing. Finally, after-ripened seed with rapid germination may be suitable for spring or summer plantings where supplemental irrigation is available.

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