Effects of revegetation on surficial soil salinity in panspot soils

D.G. HOPKINS, M.D. SWEENEY, D.R. KIRBY, AND J.L. RICHARDSON

Abstract

Panspots and transition zone pantspots (slickspots) from a Leptic/Typic Natriboroll soil complex in western North Dakota were compared to determine the effects of secondary plant succession upon soil properties. Herbage and rooting characteristics were evaluated among pantzpot, transition zone, and adjacent well-vegetated Belfield soils using point frame data and a modified dry ashing technique. The effects of vegetation upon soil electrical conductivity (EC) were tested using gradient transects aligned perpendicular to boundaries between pantzpot and transition zones. Transition zones had 40% more total forage and twice the litter found in pantzpot areas. Thirteen of 17 gradient transects showed an inverse relationship between soil EC and distance into transition zones at the 0 to 5 cm depth. Significantly higher root mass was obtained in the 0 to 5 cm depth in transition zones compared to pantzpots. A conceptual model based on surfacene water flow is presented to explain the polygonal cracking that was observed only in transition zone surfaces initiating a series of interactions resulting in natural reclamation of the transition zone soil.

Key Words: secondary succession, panspots, surface crusts, salinity

Limited productivity of rangeland vegetation on soils having natric horizons has been extensively documented (Sandoval et al. 1959, Lewis and White 1964, Stuart et al. 1971, White et al. 1982). Inhibition of vascular plant growth on panspot (slickspot) soils is especially severe due to excessive salinity and sodicity which, in effect, create "physiologic deserts" in these areas (Lewis et al. 1959, Munn and Boehm 1983).

Research on pantzpots traditionally focused on the physical and chemical characteristics of pantzpot soils as components of sodium-affected landscapes (Lewis et al. 1959, Johnson et al. 1985). Information on pantzpot vegetation has generally been limited to secondary observations made during such studies (Kellogg 1934, Mogen et al. 1959, Munn and Boehm 1983).

The identification and influence of natural vegetation dynamics on pantzpot soil properties has received little attention on the Northern Great Plains. Preliminary work in western North Dakota has indicated a potential relationship between secondary succession in pantzpots and improvements in surficial soil chemistry (Hopkins et al. 1987a).

In the field, we observed that 3 distinct range communities could be easily recognized in Natriboroll map units. Communities nearly devoid of vegetation were described as pantzpots. We noted earlier (Hopkins et al. 1987a) that soil surveyors often overestimate the extent of these soils because of their stark appearance. A second community, at the same elevation as pantzpots, supports sparse grasses and low growing half-shrubs; we refer to these areas as "transition zones". The third range community, classified as a clayey range site is found on Belfield soils. Belfield soils are at higher topographic positions than either pantzpots or transition zone soils.

This study was designed to test the hypothesis that zones of secondary plant succession in pantzpots (referred to hereafter as transition zones) modify soil electrical conductivity (EC) in the near-surface soil environment. The study objectives were to: (1) compare soil EC between pantzpot and transition zones, (2) evaluate surface and subsurface soil properties and inventory vegetation among pantzpot, transition zone, and adjacent Belfield soils, and (3) compare root density distributions between pantzpot, transition zone, and adjacent Belfield soils.

Materials and Methods

Study Area

The study area was on the Dickinson Experiment Station Ranch Headquarters approximately 40 km northwest of Dickinson, North Dakota in NE1/4, Sec. 16, T13N, R96W (Lat. 47° 12' 30"N., Long. 102° 51' 20"W.). Soils with natric horizons are common in this region of western North Dakota (Wright et al. 1982). The study area was chosen to provide detailed soil information for an ongoing short duration grazing trial in Sec. 16 (Kirby et al. 1986) and because of the high concentration of pantzpots associated with the major soils (Hopkins et al. 1987a). A fence was constructed in early June 1987 to prevent cattle from grazing the study area.

The climate of the study is continental with mean annual precipitation of 425 mm and mean air temperature of 4.6° C. About 80% of the annual precipitation falls between April and September. Mean annual snowfall is 965 mm but strong winds in the area keep most of the snow in drifts and the ground free of snow (Wright et al. 1982).

Field Methods

A 20 by 20-m area was selected that encompassed unclassified pantzpot and transition zone soils interspersed in areas of Belfield soils (fine, montmorillonitic, Glossic Natriborolls). The range site designation for the Belfield soil is clayey; pantzpot and transition zone soils are mapped as thin claypan (USDA Soil Conservation Service 1984).

Vegetation was analyzed for species composition and basal area by the point frame method (Atty and Schmid 1942) with 500 points taken in representative sites for pantzpot, transition zone, and Belfield soil areas. Five replicate 0.25-m² quadrats were clipped in similar areas to compare herbaceous and shrub standing crop among soils. Clipped samples were oven dried and weighed. Vegetation nomenclature follows the Great Plains Flora Association (1986).

Intervals of 1 m were established for detailed mapping and sampling in the 20 by 20-m study area. The soil surface was mapped and digitized to determine the extent of pantzpot, transition zone, and Belfield soils. A topographic map was made, based on 1-m observation intervals, using a surveyor's level. Slope ranges from 1 to 3% in the study area (Fig. 1). Detailed topographic data (9 points per m²) was obtained for several pantzpot areas having transition zones to determine if elevational differences were related to vegetation distribution or soil salinity in the pantzpots.

Pantzpot with transition zone areas large enough to accommodate a gradient transect were identified with the soil surface map. Seventeen transects, oriented perpendicular to the boundary between pantzpots and transition zones, were established. Sampling sites were marked off in 10-cm increments from the boundary into...
in a trench dissecting the vegetation analysis sites, using standard methods of the National Cooperative Soil Survey (Soil Survey Staff 1951). Samples were collected, air dried, and ground for laboratory characterization.

Root density distributions in panspots and transition zones were evaluated using the gradient transect method described earlier. Four root gradient transects about 1 m in length were selected near the trench and sampled with a 6.3 cm diameter coring tube to 30 cm depth to obtain sufficient volume for laboratory analysis. Soil cores were extracted at 15-cm intervals along the transects and carefully sliced in 5 cm depth increments to 15 cm, with 1 sample taken from 15 to 30 cm. Additionally, 8 sites were located randomly from the Belfield soil area to allow comparison of root density distribution among the panspot, transition zone, and Belfield soils. Root density samples were air dried for laboratory analysis.

**Laboratory Methods**

Soil electrical conductivity (EC) was determined on 1:1 soil/water suspensions for all gradient transect samples (Dahke 1980). The values for 1:1 EC are from a dilute suspension and only represent about one half of the more familiar saturated paste extract EC values. We used 1:1 suspensions because of the large number of samples and the fact that the relatively homogenous textures of the panspot soils minimize textural effects differentiating saturated paste extract EC from that of 1:1 suspensions (Hogg and Henry 1984).

Root density was determined following a method modified from Ward et al. (1978). Dry soil samples were weighed, placed in

**Table 1. Cover (%) of plant species for panspot, transition zone, and Belfield soils on the Dickinson Experiment Station Ranch Headquarters, North Dakota.**

<table>
<thead>
<tr>
<th>Species</th>
<th>Panspot</th>
<th>Transition zone</th>
<th>Belfield</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Graminoids</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Agropyron spicatum</em> (Pursh) Scrb. &amp; Smith</td>
<td>3.2</td>
<td>5.0</td>
<td>1.0</td>
</tr>
<tr>
<td><em>Agropyron caninum</em> (L.) Beauv.</td>
<td>0.4</td>
<td>---</td>
<td>0.2</td>
</tr>
<tr>
<td><em>Poa sandbergii</em>, Vasey</td>
<td>1.0</td>
<td>1.2</td>
<td>---</td>
</tr>
<tr>
<td><em>Agropyron smithii</em>, Rybd.</td>
<td>---</td>
<td>---</td>
<td>1.2</td>
</tr>
<tr>
<td><em>Stipa comata</em>, Trin. &amp; Rupr.</td>
<td>---</td>
<td>---</td>
<td>1.2</td>
</tr>
<tr>
<td><em>Stipa viridula</em>, Trin.</td>
<td>---</td>
<td>---</td>
<td>0.4</td>
</tr>
<tr>
<td><em>Koeleria pyramidata</em>, (Lam.) Beauv.</td>
<td>---</td>
<td>0.2</td>
<td>3.8</td>
</tr>
<tr>
<td><em>Bouteloua gracilis</em>, (H.B.K.) Lag. ex Griffiths</td>
<td>---</td>
<td>0.8</td>
<td>23.6</td>
</tr>
<tr>
<td><em>Schedonardus paniculatus</em> (Nutt.) Trel.</td>
<td>---</td>
<td>0.4</td>
<td>---</td>
</tr>
<tr>
<td><em>Buchloe dactyloides</em> (Nutt.) Englem.</td>
<td>---</td>
<td>---</td>
<td>2.0</td>
</tr>
<tr>
<td><em>Carex heliophila</em>, Mack.</td>
<td>---</td>
<td>---</td>
<td>2.0</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td>4.6</td>
<td>7.6</td>
<td>34.4</td>
</tr>
<tr>
<td><strong>Forbs</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Artemisia frigida</em>, Wild.</td>
<td>---</td>
<td>---</td>
<td>1.2</td>
</tr>
<tr>
<td><em>Atriplex nuttallii</em> S. Wats.</td>
<td>1.4</td>
<td>---</td>
<td>---</td>
</tr>
<tr>
<td><em>Phlox hoodii</em>, Rich.</td>
<td>---</td>
<td>---</td>
<td>1.4</td>
</tr>
<tr>
<td><em>Chenopodium glaucum</em> L.</td>
<td>0.8</td>
<td>0.4</td>
<td>---</td>
</tr>
<tr>
<td><em>Polygonum aviculare</em> L.</td>
<td>0.6</td>
<td>0.2</td>
<td>---</td>
</tr>
<tr>
<td><em>Vicia americana</em> Muhl. ex Wild.</td>
<td>---</td>
<td>0.4</td>
<td>0.4</td>
</tr>
<tr>
<td><em>Grindelia squarrosa</em> (Pursh) Dun.</td>
<td>---</td>
<td>0.8</td>
<td>---</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td>2.8</td>
<td>1.8</td>
<td>3.0</td>
</tr>
<tr>
<td><strong>Shrub</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Ceratoides lanata</em> (Pursh) Howell</td>
<td>1.0</td>
<td>1.4</td>
<td>---</td>
</tr>
<tr>
<td><em>Opuntia</em> spp.</td>
<td>0.4</td>
<td>---</td>
<td>---</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td>1.4</td>
<td>1.4</td>
<td>---</td>
</tr>
<tr>
<td><strong>Miscellaneous</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Litter</em></td>
<td>14.0</td>
<td>29.4</td>
<td>53.6</td>
</tr>
<tr>
<td><em>Bare</em></td>
<td>77.0</td>
<td>59.6</td>
<td>7.8</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td>100.0</td>
<td>100.0</td>
<td>100.0</td>
</tr>
</tbody>
</table>

1Species did not occur.

![Topographic map of study area with 1-dm contour interval, Dunn County, North Dakota; elevations in meters above MSL. Boundaries for soil characterization trench are indicated by A and B.](image-url)
fine-meshed muslin cloth and wire-wrapped. Samples were immersed overnight in a water bath with sodium hexametaphosphate to disperse soil particles. Fine soil particles were separated from the sands by gently rubbing the sample under running water. Later, the bags were opened, placed on a 100-mesh sieve, and the contents washed under running water. Roots and organic particles were separated from the sand fraction by a series of decantations. Washed root samples were placed in ashing crucibles and dried overnight at 60°C. The dried samples were cooled, weighed, and placed in an ashing oven at 470°C for 6.5 hours, then cooled and weighed.

The ashing step was included because inspection of the samples revealed that mineral matter was tightly bound to root tissues even after several shakings and decantations. To correct for tightly bound mineral matter, several mineral-free subsamples of the same root tissues were obtained and ashed. From these subsamples the average root mass lost on ashing was found to be 77.7%. The mass lost on ashing from samples containing mineral matter was divided by 0.777 to obtain a corrected dry root mass.

The 2 soils sampled in the trench were characterized in the laboratory using the following methods. Soluble salts were determined on saturated paste soil extracts using the methods of Rhoades (1982). Soil pH was determined potentiometrically (Peech 1965). Calcium, magnesium, and sodium, (Ca, Mg, and Na) were determined with atomic absorption spectroscopy using methods of Chapman and Pratt (1961). Adjusted sodium adsorption ratio (SAR_adj) was calculated using concentrations of Ca, Mg, and Na corrected for ion pairs and activities (Alzubaidi and Webster 1983) using the relationship established by the US Salinity Laboratory Staff (1954). The adjusted SAR provides a better estimate of sodium activity than SAR based on concentration alone. Organic carbon was determined using the Walkley-Black method modified by Chapman and Pratt (1961). Particle size analysis was determined by pipette method (Day 1965).

The EC data for the gradient transects were analyzed separately for each depth class using one-way analysis of variance with a covariate site along the transect (SAS Institute 1985). This approach was utilized as it provides an estimate of the effect of distance along the transect on soil EC. The presence of a functional relationship between transect site (distance) and EC for panspot and transition zone soils was observed primarily below the 793.4-m contour interval shown in Figure 1; Belfield soils were found above this contour. The distribution of the 3 soils in the study area was 13, 16, and 71% for panspot, transition zone, and Belfield soils, respectively. Panspot and transition zone soils comprise almost 90% of the landscape and present a marked contrast to well-vegetated Belfield soils.

Transition zone soils exhibit a 2-fold increase in vegetative litter compared to panspots (Table 1), but plant basal area was similar for panspot and transition zone soils (8.8 and 10.8% respectively. Table 1). There was a trend for greater yields of graminoid species in transition zones compared to panspots (Table 2). The production and diversity of species from the Belfield soil was substantially greater than from either panspot or transition zone soils (Tables 1 and 2).

Differences in basal area and forage yield mentioned above are due to differences in edaphic properties of the Belfield and panspot soils. Belfield soils had more soil organic matter to greater depths than panspot soils and lower salinity and sodicity as evidenced by EC and SAR_adj (Table 3). Depth to the C horizon of the Belfield soil was greater than in the panspot soil but both soils were underlain by highly saline siltstone.

Table 3. Selected soil morphologic, physical, and chemical properties of major soils of the study area. EC data represent about one half of saturated paste extract EC.

<table>
<thead>
<tr>
<th>Depth USDA texture2</th>
<th>Organic matter (%)</th>
<th>pH</th>
<th>EC*</th>
<th>SAR_adj</th>
</tr>
</thead>
<tbody>
<tr>
<td>Crust</td>
<td>Sil</td>
<td>1.5</td>
<td>7.5</td>
<td>2.8</td>
</tr>
<tr>
<td>E</td>
<td>Sil</td>
<td>1.8</td>
<td>7.4</td>
<td>6.0</td>
</tr>
<tr>
<td>Bt</td>
<td>Sil</td>
<td>1.6</td>
<td>7.7</td>
<td>15.6</td>
</tr>
<tr>
<td>Cl</td>
<td>SilCL</td>
<td>1.1</td>
<td>7.8</td>
<td>17.7</td>
</tr>
<tr>
<td>C2</td>
<td>Sil</td>
<td>0.7</td>
<td>7.7</td>
<td>16.7</td>
</tr>
<tr>
<td>C1</td>
<td>Sil</td>
<td>0.3</td>
<td>7.8</td>
<td>16.0</td>
</tr>
<tr>
<td>C2</td>
<td>Sil</td>
<td>0.3</td>
<td>7.4</td>
<td>15.7</td>
</tr>
<tr>
<td>C3</td>
<td>Sil</td>
<td>0.4</td>
<td>7.4</td>
<td>13.0</td>
</tr>
<tr>
<td>Bt1</td>
<td>Sil</td>
<td>6.6</td>
<td>7.3</td>
<td>2.0</td>
</tr>
<tr>
<td>Bt2</td>
<td>SilCL</td>
<td>2.5</td>
<td>7.2</td>
<td>0.4</td>
</tr>
<tr>
<td>Bt1</td>
<td>Sil</td>
<td>1.3</td>
<td>7.8</td>
<td>0.8</td>
</tr>
<tr>
<td>C</td>
<td>Sil</td>
<td>1.4</td>
<td>8.1</td>
<td>0.7</td>
</tr>
<tr>
<td>C1</td>
<td>SilCL</td>
<td>1.1</td>
<td>8.0</td>
<td>4.8</td>
</tr>
<tr>
<td>C2</td>
<td>Sil</td>
<td>0.7</td>
<td>8.2</td>
<td>16.0</td>
</tr>
<tr>
<td>C2</td>
<td>Sil</td>
<td>0.6</td>
<td>7.9</td>
<td>14.9</td>
</tr>
</tbody>
</table>

1 nomenclature follows USDA/SCS standards
2 Si-silt, L-loam, C-clay
3 1:1 soil/water suspension

Variations in salinity were large in the locality of the study area and even larger in panspot soils (Hopkins et al. 1987a). We generalized the data on salinity variability (as measured by EC) by pooling observations from all sampling sites and transects (Table 4). These data were stratified by depth class. Mean EC increases with depth, and standard errors usually decrease with depth.

The relationship between soil EC and site (distance on transects) was not consistent for transects in the study area (data not shown). Due to the presence of significant interaction, individual regression coefficients were calculated for each transect (Table 5). Soil EC and transect site were inversely related for the 0 to 3 cm depth class in 13 of 17 transects (Table 5) indicating that soil EC was lower in transition zones. The number of transects with negative regression coefficients decreases in lower depth classes. In 4 transects there was an increase in magnitude of negative coefficients in the second compared to the first depth class.

Table 2. Forage yield of panspot, transition zones, and Belfield soils on the Dickinson Experiment Station Ranch Headquarters, North Dakota.

<table>
<thead>
<tr>
<th>Vegetation type</th>
<th>Panspot soil</th>
<th>Transition zone soil</th>
<th>Belfield soil</th>
</tr>
</thead>
<tbody>
<tr>
<td>Forb</td>
<td>15 (2.6)</td>
<td>22 (11.2)</td>
<td>87 (5.6)</td>
</tr>
<tr>
<td>Shrubs</td>
<td>30 (1.2)</td>
<td>2 (0.9)</td>
<td>20 (19.2)</td>
</tr>
<tr>
<td>Total</td>
<td>6 (7.5)</td>
<td>48 (7.5)</td>
<td>6 (2.6)</td>
</tr>
</tbody>
</table>

1 Standard error.
had only slightly higher values in the transition zone with a maximum found on the boundary.

**Discussion**

**Vegetation Analyses**

Forage yield, litter cover, and species composition vary markedly for panspot, transition zone, and Belfield soils. The distinctive range communities are in close proximity and are characteristic of sodium-affected landscapes (Sandoval and Reichman 1971, White et al. 1981). Salt tolerant species identified on panspot and transition zone soils of this study were widely recognized on other sodium-affected soils of the Northern Great Plains and in soils in big sagebrush communities (Artemisia tridentata tridentata Nutt.) of the Great Basin Physiographic Province (Hugie and Passey 1964; Eckert et al. 1978, 1986).

Increased litter in transition zone areas creates higher albedo, which has been reported to affect substantial changes in both the thermal regime of soils and the resultant soil moisture status (Oke 1978). Litter also acts as a mulch, reducing the amount of moisture lost from the soil by damping maximum and minimum soil surface air temperatures and increasing humidity. The mulch effect was demonstrated by Evans and Young (1970) in their study on population dynamics in cheatgrass (Bromus tectorum L.). These microsite properties may maintain a higher humidity and a more moderate thermal regime in transition zones as compared to panspot soils.

Cryptogams were observed to be an integral vegetative component in transition zone soils (Fig. 3). *Nostoc* spp. were the

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**Fig. 2.** Rootmass comparisons (as percent of dry soil weight) between transects 3 and 4 at 0–5 cm depth as a function of distance on transect.

**Fig. 3.** Cryptogamic vegetation (*Nostoc* spp.) in a transition zone (note polygonal cracking).

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**Table 4. Variations in soil salinity of panspot and transition zone soils of the study area pooled for all transects by depth class.**

<table>
<thead>
<tr>
<th>Depth midpoints (cm)</th>
<th>Mean EC dS m⁻¹</th>
<th>Standard Deviation</th>
<th>Standard error</th>
</tr>
</thead>
<tbody>
<tr>
<td>2.5</td>
<td>1.98</td>
<td>0.56</td>
<td>0.14</td>
</tr>
<tr>
<td>7.5</td>
<td>5.39</td>
<td>1.33</td>
<td>0.32</td>
</tr>
<tr>
<td>12.5</td>
<td>7.54</td>
<td>1.04</td>
<td>0.25</td>
</tr>
<tr>
<td>17.5</td>
<td>8.59</td>
<td>0.72</td>
<td>0.18</td>
</tr>
<tr>
<td>22.5</td>
<td>9.26</td>
<td>0.36</td>
<td>0.09</td>
</tr>
<tr>
<td>27.5</td>
<td>9.62</td>
<td>0.41</td>
<td>0.10</td>
</tr>
</tbody>
</table>

---

**Table 5. Sample regression coefficients (b) for all gradient transects by depth class.**

<table>
<thead>
<tr>
<th>Transect</th>
<th>Depth midpoints (cm)</th>
<th>n</th>
<th>2.5</th>
<th>7.5</th>
<th>12.5</th>
<th>17.5</th>
<th>22.5</th>
<th>27.5</th>
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<tr>
<td></td>
<td></td>
<td>1</td>
<td>-0.025</td>
<td>-0.022</td>
<td>-0.049*</td>
<td>-0.035*</td>
<td>-0.050*</td>
<td>-0.026</td>
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<tr>
<td></td>
<td></td>
<td>2</td>
<td>-0.048*</td>
<td>-0.021</td>
<td>-0.005</td>
<td>0.000</td>
<td>0.004</td>
<td>0.028</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3</td>
<td>-0.035*</td>
<td>-0.058*</td>
<td>-0.040*</td>
<td>-0.026*</td>
<td>-0.023*</td>
<td>-0.020*</td>
</tr>
<tr>
<td></td>
<td></td>
<td>4</td>
<td>0.005</td>
<td>0.007</td>
<td>-0.003</td>
<td>-0.012*</td>
<td>-0.017#</td>
<td>0.018*</td>
</tr>
<tr>
<td></td>
<td></td>
<td>5</td>
<td>-0.002</td>
<td>0.009</td>
<td>0.031*</td>
<td>0.014*</td>
<td>0.021*</td>
<td>0.028*</td>
</tr>
<tr>
<td></td>
<td></td>
<td>6</td>
<td>0.005</td>
<td>-0.040#</td>
<td>-0.034#</td>
<td>-0.020*</td>
<td>-0.007</td>
<td>0.000</td>
</tr>
<tr>
<td></td>
<td></td>
<td>7</td>
<td>-0.003</td>
<td>0.020*</td>
<td>0.014*</td>
<td>0.026*</td>
<td>0.024*</td>
<td>0.015*</td>
</tr>
<tr>
<td></td>
<td></td>
<td>8</td>
<td>0.006</td>
<td>0.046*</td>
<td>0.045*</td>
<td>0.009</td>
<td>0.029</td>
<td>-0.024</td>
</tr>
<tr>
<td></td>
<td></td>
<td>9</td>
<td>-0.013</td>
<td>-0.014</td>
<td>0.000</td>
<td>0.002</td>
<td>-0.003</td>
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<td></td>
<td></td>
<td>10</td>
<td>-0.005</td>
<td>-0.005</td>
<td>0.001</td>
<td>-0.001</td>
<td>-0.002</td>
<td>0.019*</td>
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<tr>
<td></td>
<td></td>
<td>11</td>
<td>-0.070*</td>
<td>-0.100#</td>
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<td>-0.025</td>
<td>-0.035</td>
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<td>-0.010</td>
<td>0.050</td>
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<td>-0.065</td>
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<tr>
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<td>-0.077*</td>
<td>-0.007</td>
<td>0.020</td>
<td>-0.005</td>
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<tr>
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<td>-0.026</td>
<td>-0.028</td>
<td>-0.009</td>
<td>0.009</td>
<td>-0.007</td>
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<tr>
<td></td>
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<td>15</td>
<td>-0.005</td>
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<td>-0.025</td>
<td>0.030</td>
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<tr>
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<td>-0.093*</td>
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<td>-0.046*</td>
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<td>0.007</td>
<td>-0.001</td>
<td>-0.001</td>
<td>-0.010</td>
<td>0.002</td>
</tr>
</tbody>
</table>

Regression coefficients for transects 5, 7, and 8 deviated from the other transects; soil EC increases with distance into the transition zone at most depths. These transects appear responsible for the significant site-transect interaction in the overall analysis.

Mean rootmass for the 0–30 cm depth is 0.17, 0.29, and 1.39%, respectively, for panspot, transition zone, and Belfield soils. Root density distributions were greater for all depth classes in transition zones compared to panspots but only the 0 to 5 cm depth showed significantly higher rootmass in transition zone soils (Table 6).

**Table 6. Rootmass means (% dry soil weight), standard deviations, and standard errors by individual and pooled depth classes for panspots (pan) and transition zone (transition) soils.**

<table>
<thead>
<tr>
<th>Depth class (cm)</th>
<th>Mean Pan</th>
<th>Standard Dev. Pan</th>
<th>Standard error Pan</th>
<th>Mean Transition</th>
<th>Standard Dev. Transition</th>
<th>Standard error Transition</th>
</tr>
</thead>
<tbody>
<tr>
<td>0–30</td>
<td>0.29</td>
<td>0.08</td>
<td>0.08</td>
<td>0.04</td>
<td>0.04</td>
<td>0.07</td>
</tr>
</tbody>
</table>

Incremental increases in rootmass with distance along transects were inconsistent (Fig. 2). Transect 3 illustrates a gradual increase in rootmass with distance into the transition zone, but transect 4
dominant cryptogam (T. Esslinger, personal communication). Cryptogams have been extensively studied in the Great Basin Physiographic Province; Anderson et al. (1982) state that the success of algae and fungi on saline sites is "undoubtedly related to physiologic tolerance." The presence of algal species has been noted locally in panspot soils by White et al. (1981). In this study algal growth seems adapted to transition zone areas; little algae was observed in panspots.

Polygonal cracking was observed in transition zone soils in preliminary fieldwork (Hopkins et al. 1987b), and close inspection of the landscape during this study revealed that polygonal cracking was restricted to transition zone soils. Soil microtopography, especially cracking, was shown by Harper et al. (1965) to be a major factor controlling seed germination, because microtopographic variations "determine the frequency on the soil surface of microsites sufficiently humid for germination". In an extensive sampling of natural vegetation on crusted Aridisols in Nevada, Eckert et al. (1986) observed greater plant densities in trenches and polygonal cracks than on adjacent crusted soils.

**Root Density Distribution**

We believe the transition zones are not ephemeral phenomenon as evidenced by significantly higher rootmass values in transition zones compared to panspots for the 0 to 5 cm depth. Increased rooting activity results in increased organic matter contents and available water capacities in transition zone soil surfaces. Belfield soils are markedly different from panspot and transition zone soils and exhibit a 4-fold increase in root biomass in the 0 to 30 cm depth.

The presence of roots in panspots poses a question of origin. There is a possibility that rootmass values reflect sporadic growth of halophytic vegetation in moist years. The rare occasion of moisture in panspots when biotic activity is high, however, suggests that organic matter decomposition would be minimal. Alternatively, these roots may be relict features. Johnson et al. (1985) have shown that panspots are "erosional features that tend to move upslope". The panspots in this study are similar features; roots found in these panspots may have developed under different soil conditions and exist in a somewhat preserved state.

**Gradient Transect Analysis**

The number of transects with negative regression coefficients in the 0 to 5 cm depth class indicates a strong trend supporting our hypothesis of reduced soil EC in transition zones (Table 5). Vegetation establishment and growth in transition zones may increase infiltration and slowly improve soil structure resulting in lower levels of soil salinity. Supporting evidence for this process was reported by Hopkins et al. (1990). Significant reductions for both depth to gypsum and weighted soil EC in transition zones were obtained from soil pedons sampled uniformly on this study site. Vegetation establishment in transition zones, however, may simply reflect random distribution of zones of lower soil salinity in panspot soils; seeds are preferentially established where osmotic stress is minimized.

Lewis et al. (1959) used radioactive sulfur to trace soil moisture movement in panspot soils and demonstrated that "soil moisture moved into the panspot soil at some undetermined depth from the surrounding associated soil". The gradient for water flow was caused by the very low osmotic/matrix potential for the panspot soil compared to adjacent soils characterized by relatively rapid, downward water flow. These adjacent soils, similar to Belfield soils in our study, act essentially as recharge zones. Water is transferred through the solum to saline, impermeable substrata, and becomes concentrated, following the water potential gradient to panspots, which are the hydraulic discharge zones. As the discharge zones dry, polygonal cracks form due to the high percentage of smectite.

The polygonal cracking that we observed in transition zone soils does not appear to be caused by elevational gradients between panspots and transition zones. Results from comparisons of microtopographic elevation indicate less than 5 mm difference between panspot and transition zone surfaces. In many cases transition zones were the same elevation as adjacent panspot surfaces. Differential ponding following precipitation does not appear to be a factor in development of polygonal cracking or reductions in soil salinity in transition zones.

Continued maintenance of plant growth in transition zones fosters the beneficial properties mentioned earlier and effects the reversal of water flow in transition zones from discharge to recharge. The initiation of a leaching profile in transition zones is the first step towards natural reclamation of the panspot/transition soil complex.

**Literature Cited**


