Research Note

Effect of Types of Biosolids and Cattle Manure on Desert Grass Growth

Ricardo Mata-González,¹ Ronald E. Sosebee,² and Changgui Wan³

Authors are ¹Ecologist, MWH Americas, Inc, Fort Collins, CO 80525; and ²Professor and ³Research Associate, Department of Range, Wildlife, and Fisheries Management, Texas Tech University, Lubbock, TX 79409.

Abstract

We compared the effect of applying anaerobically produced biosolids, lime-stabilized biosolids, and cattle manure on the production of blue grama (*Bouteloua gracilis* [HBK] Lag. ex Steud.) and black grama (*B. eriopoda* [Torr.] Torr.) grown in pots with moderate soil water content. We also compared the physicochemical and bacteriological composition of these 3 amendments. All amendments produced similar increases in plant growth, despite their differences in plant nutrient concentrations, as a result of limitations in soil water. Heavy metal levels in biosolids were within the US Environmental Protection agency limits for Class A and Class B biosolids, but were higher than in manure. In contrast, pathogen levels were lower in biosolids than in manure. Application of biosolids and cattle manure increased the production of both grasses and may have equivalent effects under typical climatic and soil conditions of semiarid rangelands.

Resumen

Comparamos el efecto de la aplicación de biosólidos producidos anaeróbicamente, biosólidos producidos por aplicación de cal, y estiércol bovino en la producción de navajita azul (*Bouteloua gracilis* [HBK] Lag. ex Steud.) y navajita negro (*B. eriopoda* [Torr.] Torr.) creciendo en macetas con nivel moderado de humedad. También comparamos la composición físico-química y bacteriológica de estos tres productos orgánicos. Los tres productos orgánicos produjeron un incremento similar en el crecimiento de plants a pesar de sus diferencias en niveles de nutrientes, debido a limitaciones en humedad. Los metales pesados en biosólidos estuvieron dentro de los límites establecidos por la US Environmental Protection Agency para biosólidos que en estiércol. La aplicacion de biosólidos y estiércol incremento la producción de los dos pastos y pueden tener efectos equivalentes bajo condiciones climáticas y edáficas de pastizales semiáridos.

Key Words: black grama, blue grama, Bouteloua eriopoda, Bouteloua gracilis, fecal coliform bacteria, heavy metals

INTRODUCTION

Biosolids are nutrient-rich organic compounds that result as a by-product of municipal wastewater treatment. The beneficial use of biosolids as soil amendment and the enhancement of rangeland plant production have been documented (Fresquez et al. 1990; Jurado and Wester 2001; Mata-González et al. 2002, 2004). Biosolids, however, require certain quality standards to prevent contamination. Class B biosolids, with low levels of pathogens and heavy metals, are required for application in rangelands, agricultural lands, and reclamation sites. Class A biosolids, with even lower levels of pathogens and heavy metals, are required for application in lawns or home gardens (USEPA 1994, Table 1). Different municipal treatment methods may affect the composition and the fertilizing characteristics of the biosolids (USEPA 2000). Many studies on rangeland plants have used biosolids obtained by anaerobic digestion (Fresquez et al. 1990; Jurado and Wester 2001; Mata-González et al. 2002, 2004). Biosolids obtained by lime stabilization have rarely been used in studies despite their abundance (USEPA 2000).

Biosolids commonly have total N content of 3%–5%, which is comparable to animal manures (Ajwa and Tabatabai 1994). Although their fertilizing effect could be similar, there is limited information (Moss et al. 2002) directly comparing the benefits of biosolids and manure on rangeland plant growth. Such information is important to associate the effect of biosolids to that of the more conventional and socially accepted cattle manure (Moss et al. 2002).

In this study we compared the effect of applying anaerobically produced biosolids, lime-stabilized biosolids, and cattle manure as soil amendments on aboveground production of blue grama (*Bouteloua gracilis* [HBK] Lag. ex Steud.) and black grama (*Bouteloua eriopoda* [Torr.] Torr.), grown in pots within a rainout shelter. Our hypothesis was that the 3 amendments would produce similar effects on desert grass growth. We also compared the amendments in terms of some physicochemical

Biosolids and partial funding were provided by MERCO Joint Venture LLP. Assistance from Pedro Jurado Guerra, David Wester, Carlos Villalobos, Corey Moffet, and Joanna Hahm is gratefully acknowledged. This is contribution T-9-1051 of the College of Agricultural Sciences and Natural Resources, Texas Tech University.

Correspondence: R. Mata-González, MWH Americas, Inc, 1825 Sharp Point Drive, Suite 118, Fort Collins, CO 80525. Email: ricardo.mata-gonzalez@mwhglobal.com

Manuscript received 5 October 2005; manuscript accepted 31 August 2006.

Table	1. Physico	chemical and	bacteriologica	I componen	its (mean	\pm SEM) of 2 types	of bioso	olids and	cattle	manure	and US	SEPA (199	94) limi	its for
heavy	metals and	pathogens in	Class A and	B biosolids.	Different	letters i	ndicate ame	ndment	difference	es (P	< 0.05) for a	particular	compo	onent.

	Anaerobically	Lime-stabilized		USEPA limits	USEPA limits
Component	produced biosolids	biosolids	Cattle manure	for class B	for class A
$TKN^1 (g \cdot kg^{-1})$	49.2 ± 2.2 a	50.6 ± 1.8 a	12.1 \pm 0.5 b	N/A	N/A
$P(g \cdot kg^{-1})$	1.6 \pm 0.1 a	1.4 ± 0.1 a	0.3 ± 0.1 b	N/A	N/A
$K (g \cdot kg^{-1})$	1.6 \pm 0.1 a	1.4 ± 0.1 a	4.2 ± 0.3 b	N/A	N/A
Ca $(g \cdot kg^{-1})$	31.4 \pm 2.3 a	$41.6\pm4.0~\text{b}$	$19.0\pm0.7~c$	N/A	N/A
Mg $(g \cdot kg^{-1})$	7.7 ± 0.2 a	7.1 ± 0.3 a	2.7 ± 0.1 b	N/A	N/A
$S(g \cdot kg^{-1})$	16.0 \pm 0.5 a	17.6 ± 1.1 a	2.0 ± 0.1 b	N/A	N/A
Na (g•kg ⁻¹)	1.7 ± 0.4 a	$2.6~\pm~0.4$ b	2.3 ± 0.1 b	N/A	N/A
EC ($dS \cdot m^{-1}$)	9.9 ± 0.9 a	12.5 \pm 0.9 b	$4.3\pm0.3~\text{c}$	N/A	N/A
pН	7.3 ± 0.1 a	7.3 ± 0.1 a	6.9 ± 0.1 b	N/A	N/A
Water $(g \cdot kg^{-1})$	798.1 ± 3.0 a	802.0 ± 3.0 a	711.8 \pm 8.7 b	N/A	N/A
$Mn (mg kg^{-1})$	573.0 ± 7.4 a	567.0 ± 22.6 a	133.6 \pm 4.5 b	N/A	N/A
Ni (mg·kg ⁻¹)	$44.4~\pm~3.3~a$	54.5 \pm 4.0 b	6.0 ± 0.3 c	420	420
Cd (mg \cdot kg ⁻¹)	3.3 ± 0.2 a	3.6 ± 0.3 a	$3.6~\pm~3.4$ a	85	39
Zn (mg \cdot kg ⁻¹)	$1258 \pm 48.3 a$	$1105 \pm 65.7 a$	$84~\pm~14.8$ b	7 500	2 800
Pb (mg \cdot kg ⁻¹)	$221.2 \pm 3.5 a$	$211.8\pm9.4~a$	7.3 ± 0.8 b	840	300
Cu (mg·kg ⁻¹)	920.6 \pm 48.9 a	762.0 \pm 53.4 b	$25.2\pm2.6~\text{c}$	4 300	1 500
Fecal coliform bacteria					
$(CFU \cdot g^{-1})$	$< 1 x 10^{6} a$	$< 1 \text{ x } 10^{6} \text{ a}$	$> 50 \text{ x } 10^6 \text{ b}$	$< 2 \text{ x } 10^{6}$	< 1 000

¹TKN indicates total Kjeldahl nitrogen; EC, electrical conductivity; CFU, colony-forming units.

and bacteriological components and their potential benefits and disadvantages for land application.

MATERIALS AND METHODS

This study was conducted at the Sierra Blanca Ranch, Texas, where blue grama and black grama are important components of desert grasslands. This area is part of the Chihuahuan Desert, with annual precipitation of 310 mm concentrated from July to September. Plants of both species with 4 tillers were collected from the field in May 2000 and transplanted into plastic pots (volume 14.7 l, depth 30 cm, and diameter 25 cm) containing Armesa taxadjunct fine sandy loam soil, which is a fine, loamy, mixed, thermic Ustic Haplocalcid. This soil holds about 190 g of water per kg of soil at field capacity, has a pH of 7.5, a TKN level of $496 \text{ mg} \cdot \text{kg}^{-1}$, and an organic matter content of 7 $g \cdot kg^{-1}$ (more details in Mata-González et al. 2004). The soil used in the experiment was collected from 0-30 cm depth, air dried, and passed through a 0.5-cm sieve to provide a homogeneous rooting medium and to facilitate water distribution in the pot.

The plants were grown under a semicylindrical rainout shelter covered with greenhouse plastic film. The plants were regularly irrigated to achieve establishment for 2 months before initiating the biosolids treatments. Anaerobically produced biosolids, lime-stabilized biosolids, and cattle manure were applied on the soil surface of the pots in early July 2000 at rates of 0 (control), 34, and 90 dry Mg·ha⁻¹. The biosolids rate of 90 Mg·ha⁻¹ was selected for research purposes only since it is considered an excessive rate for commercial purposes (Mata-González et al. 2002). Biosolids were of residential origin from New York City. All biosolids were initially produced by anaerobic digestion, but some batches that did not qualify as Class B were additionally treated (lime-stabilized) with quicklime (CaO) at the Sierra Blanca Ranch to achieve Class B quality. Cattle manure was collected fresh from steers grazing in native pastures on the Sierra Blanca Ranch.

Chemical and bacteriological analyses of biosolids and cattle manure were estimated from 5 samples that were taken immediately after the application to the experimental pots. These analyses were made at the Soil, Water, and Air Testing (SWAT) laboratory of New Mexico State University, Las Cruces, New Mexico. Biosolids TKN was analyzed by the macro Kjeldahl method. The concentrations of P, K, Ca, Mg, S, Na, and heavy metals were obtained using inductively coupled plasma-atomic emission spectrometry (EPA Method 200.7) (USEPA 2001). Electrical conductivity and pH were determined in a solution extracted from a saturated paste. The bacteriological determination consisted of counts of colony forming units of fecal coliform bacteria by the membrane filter procedure (SM 9222D) (APHA 1998).

The experimental unit was a pot containing 1 plant of either blue grama or black grama. A factorial experiment (3×3) was established by combining 3 organic amendments: anaerobically produced biosolids, lime-stabilized biosolids, and cattle manure at 3 application rates: 0, 34, and 90 dry Mg·ha⁻¹. As a result, 9 treatments were applied to each species, and each experimental unit was replicated 4 times. The pots were arranged in a completely randomized design within the rainout shelter.

After the application of biosolids and cattle manure, pots were homogeneously watered at 60% of field capacity. Previous experience (Mata-González et al. 2004) with the same soil guided to select this irrigation level, achieving a moderate and consistent soil water content. Applying moderate levels of water had the objective of more accurately representing the predominant dry conditions of the study area. The pots were weekly weighed and irrigated with the necessary amount of water to restore the soil water level. The treatments were maintained for 3 months, after which plants were harvested at ground level, oven dried at 75°C for 24 hours, and weighed. Data were analyzed by analysis of variance, and significant differences among treatments were determined at P < 0.05 by the protected LSD test.

RESULTS AND DISCUSSION

There was no interaction between amendments and application rates for either blue grama (P = 0.91) or black grama (P = 0.10). Therefore, the effects of these 2 factors were independent. There were no differences in aboveground biomass production of the 2 grasses (P = 0.90) among the 3 organic amendments (Fig. 1). This suggests that no differences in grass growth can be expected by applying any of these amendments to desert grasslands under similar soil water conditions. The levels of N, P, and K were very similar in both types of biosolids (Table 1), indicating that their fertilizing effect would be comparable. Cattle manure, however, had only one-fourth of the N and one-sixth of the P contained in the biosolids and yet had the same effect as biosolids on plant growth. An important factor that may have influenced this response was the relatively low soil moisture level (60% field capacity) at which the plants were maintained. A previous study (Mata-González et al. 2004) reported that the fertilizing effect of biosolids was severely limited by low soil water content (40% field capacity). Stavast et al. (2005) also reported the effect of manure application to blue grama was restricted



Figure 1. Aboveground dry weight per plant (mean \pm SEM) of blue grama and black grama as affected by application of different types of biosolids and cattle manure irrespective of the rate of application.

by low rainfall. In our study both grasses were apparently water limited and thus unable to benefit from the higher levels of nutrients of biosolids in comparison to manure. Although our study cannot accurately represent field conditions, results suggest that under water-limiting conditions, applying biosolids and manure with different nutrient concentrations would produce similar effects in desert grass production.

Regardless of the type, the application of organic amendments increased aboveground production of both grasses with respect to the control (Fig. 2). Application of 34 Mg \cdot ha⁻¹ was marginally higher (P = 0.07) than the control, while application of 90 Mg \cdot ha⁻¹ was higher (P = 0.02) than the other 2



Figure 2. Aboveground dry weight per plant (mean \pm SEM) of blue grama and black grama as affected by application of different rates of organic amendments irrespective of the type of amendment.

application rates. Aboveground desert grass growth has been previously documented following application of biosolids (Fresquez et al. 1990; Jurado and Wester 2001; Mata-González et al. 2002, 2004) and cattle manure (Stavast et al. 2005), but direct comparisons of these 2 amendments have been lacking (Moss et al. 2002). Root growth was not evaluated in this study, but a previous report (Mata-González et al. 2002) showed that biosolids tend to favor shoot over root production.

Blue grama has frequently been the subject of biosolids and cattle manure studies (Fresquez et al. 1990; Mata-González et al. 2002, 2004; Stavast et al. 2005), but this is the first report of black grama. Our results showed that blue grama and black grama responded similarly to the application of organic amendments, despite indications that black grama could be more responsive to environmental variation than blue grama (Gosz and Gosz 1996).

Despite the benefit of higher levels of N and P with respect to manure, biosolids had the disadvantage of higher levels of electrical conductivity and heavy metals (although Cd was equally present in biosolids and manure) (Table 1). Nevertheless, the heavy metal concentrations in both types of biosolids were well within the standards for biosolids of Class A and B (USEPA 1994). Complying with these standards and particularly applying biosolids in arid areas, where low moisture and high pH restrict heavy metals mobility, contamination of soils and plants is highly unlikely (Fresquez et al. 1990). The application of 90 Mg·ha⁻¹, however, could represent an unacceptable risk (Fresquez et al. 1990). As previously stated, this rate was selected for research purposes only, and it is not recommended for commercial applications (Mata-González et al. 2002).

Cattle manure had much higher levels of fecal coliform bacteria than biosolids (Table 1). In effect, following the USEPA (1994) bacteriological standards for biosolids application, our manure samples would not qualify for land application. However, the levels of pathogens in manure are not strictly regulated as biosolids are, and yet land application of manure is more common and more socially accepted than application of biosolids (Moss et al. 2002). Likely, the public perception toward biosolids application will tend to be more favorable as the safe beneficial use of biosolids increases.

MANAGEMENT IMPLICATIONS

In summary, our results suggest that in terms of desert grass production, applying anaerobically produced biosolids, limestabilized biosolids, and cattle manure may have equivalent beneficial effects. In addition, applying biosolids could be, in bacteriological terms, even safer for human health than applying manure. Our results also warrant field investigation as a step forward in comparing the effect of biosolids and manure on rangelands. Although pot studies provide welldefined environments in which plants grow, conclusions are limited by the transferability of results to field conditions, with much higher environmental variation.

LITERATURE CITED

- AJWA, H. A., AND M. A. TABATABAI. 1994. Decomposition of different organic materials in soils. *Biology and Fertility of Soils* 18:175–182.
- APHA (AMERICAN PUBLIC HEALTH ASSOCIATION). 1998. Standard methods for the examination of water and wastewater. 20th ed. Washington, DC: American Public Health Association.
- FRESQUEZ, P. R., R. E. FRANCIS, AND G. L. DENNIS. 1990. Sewage sludge effects on soil and plant quality in a degraded, semiarid grassland. *Journal of Environmental Quality* 19:324–329.
- Gosz, R. J., AND J. R. Gosz. 1996. Species interactions on the biome transition zone in New Mexico: response of blue grama (*Bouteloua gracilis*) and black grama (*Bouteloua eriopoda*) to fire and herbivory. *Journal of Arid Environments* 34:101–114.
- JURADO, P., AND D. B. WESTER. 2001. Effects of biosolids on tobosagrass growth in the Chihuahuan desert. *Journal of Range Management* 54:89–95.
- MATA-GONZÁLEZ, R., R. E. SOSEBEE, AND C. WAN. 2002. Shoot and root biomass of desert grasses as affected by application of biosolids. *Journal of Arid Environments* 50:477–488.
- MATA-GONZÁLEZ, R., R. E. SOSEBEE, AND C. WAN. 2004. Nitrogen in desert grasses as affected by biosolids, their time of application, and soil water content. *Arid Land Research and Management* 18:385–395.
- Moss, L. H., E. EPSTEIN, AND T. L. LOGAN. 2002. Evaluating risks and benefits of soil amendments used in agriculture. Report 99-PUM-1. Alexandria, VA: Water Environment Research Foundation.
- STAVAST, L. J., T. T. BAKER, A. L. ULERY, R. P. FLYNN, M. K. WOOD, AND D. S. CRAM. 2005. New Mexico blue grama rangeland response to daily manure application. *Rangeland Ecology & Management* 58:423–429.
- USEPA (UNITED STATES ENVIRONMENTAL PROTECTION AGENCY). 1994. A plain English guide to the EPA part 503 biosolids rule. EPA832/r-93/003. Washington, DC: US Government Printing Office.
- USEPA (UNITED STATES ENVIRONMENTAL PROTECTION AGENCY). 2000. Alkaline stabilization of biosolids. Biosolids technology fact sheet. EPA 832-F-00-052. Washington, DC: US Government Printing Office.
- USEPA (UNITED STATES ENVIRONMENTAL PROTECTION AGENCY). 2001. Method 200.7, trace elements in water, solids, and biosolids by inductively coupled plasma-atomic emission spectrometry. EPA-821-R-01-010. Washington, DC: US Government Printing Office.